



**National
Oceanography Centre**
NATURAL ENVIRONMENT RESEARCH COUNCIL

National Oceanography Centre

Cruise Report No. 49

RRS *Discovery* Cruise DY026

03 - 24 AUG 2014

Shelf seas biogeochemistry cruise to the Celtic Sea

Principal Scientists

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2017

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<i>ABSTRACT</i> <p>DY26a formed part of the NERC shelf sea biogeochemistry programme. It had four principal objectives:</p> <ol style="list-style-type: none">1. To continue the seasonal time series sampling at the three key sites (Shelf break, Candyfloss, Celtic Deep).2. To provide sampling opportunities for the Shelf sea biogeochemistry students3. To obtain samples of sinking particulate organic matter at differing stages of the tidal cycle to examine the role of tidal resuspension on elemental cycling beneath the thermocline.4. To trial autonomous nitrate sensors CTDs and gliders as part of the Sensors on Gliders programme <p>The main objective of DY026b was to service the moorings. Specifically:</p> <ol style="list-style-type: none">a) To service 8 moorings/landers distributed across 5 different sites in the Celtic Seab) To calibrate the moorings	
<i>KEYWORDS</i>	
<i>ISSUING ORGANISATION</i> National Oceanography Centre University of Southampton Waterfront Campus European Way Southampton SO14 3ZH UK Tel: +44(0)23 80596116 Email: nol@noc.soton.ac.uk <i>A pdf of this report is available for download at: http://eprints.soton.ac.uk</i>	

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Scientific Personnel DY026a

Sanders, Richard	(Principal Scientist)	National Oceanography Centre Southampton
Barnett, Michelle	(MSc Student)	University of Southampton
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Cavan, Emma	(PhD Student)	University of Southampton
Clark, Darren		Plymouth Marine Laboratory
Davis, Clare		University of Liverpool
Garcia-Martin, E Elena		University of East Anglia
Giering, Sarah L C		National Oceanography Centre Southampton
Mahaffey, Claire		University of Liverpool
McNeil, Sharon		Scottish Association for Marine Science
Seguro Chata, M Isabel		University of East Anglia
Short, Jon	(Project Manager NMF)	National Oceanography Centre Southampton
Sims, Richard		Plymouth Marine Laboratory
Walk, John	(Sensors Group OTE)	National Oceanography Centre Southampton
Ward, Samuel	(MARS)	National Oceanography Centre Southampton
Woodward, Malcolm		Plymouth Marine Laboratory
Woodward, Stephen	(MARS)	National Oceanography Centre Southampton

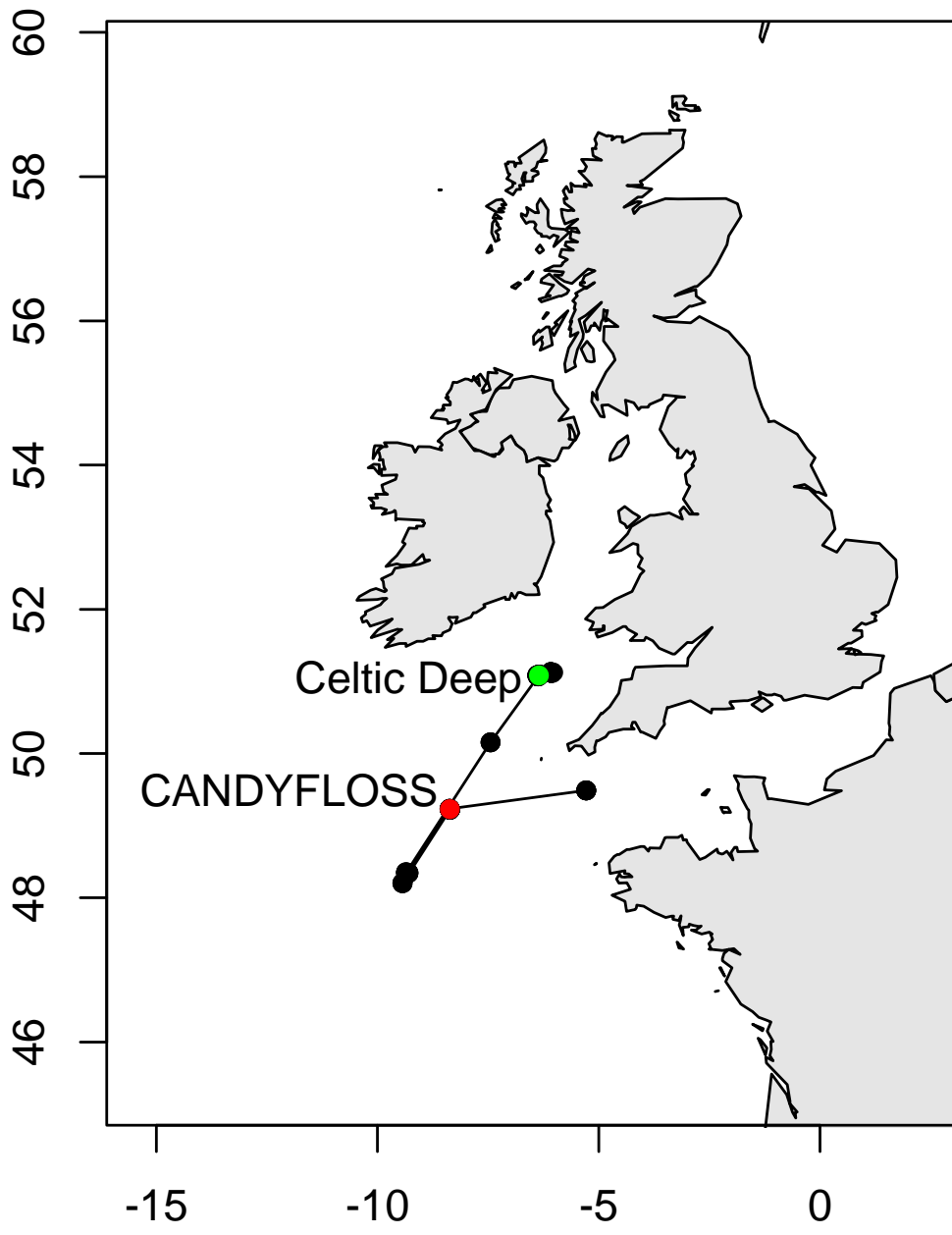
Scientific Personnel DY026b

Sivyer, David	(Principal Scientist)	CEFAS
Curran, Kieran		Plymouth Marine Laboratory
Fox, James		University of Essex
Hartman, Susan		National Oceanography Centre Southampton
Hopkins, Joanne		National Oceanography Centre, Liverpool
Mahaffey, Claire		University of Liverpool
Painter, Stuart		National Oceanography Centre Southampton
Palmer, Matthew		National Oceanography Centre Liverpool
Poulton, Alex		National Oceanography Centre Southampton
Rippeth, Tom		University of Bangor
Souza, Alex		National Oceanography Centre, Liverpool
Statham, Peter		University of Southampton
Ward, Samuel	(MARS)	National Oceanography Centre, Southampton
White, David	(MARS)	National Oceanography Centre, Southampton
Woodward, Malcolm		Plymouth Marine Laboratory

Ship's Personnel

Gwinnell, James	Master
Warner, Richard Alan	Chief Officer
Laidlow, Vanessa	2 nd Chief Officer
Tulloch, Daniel	3 rd Chief Officer
Gorbacovs, Aleksandrs	Chief Engineer
Hagan, John Andrew	2 nd Engineer
Nadkar, Simon Vivek	3 rd Engineer
Silajdziv, Edin	3 rd Engineer
Appleton, Philip Kevin	ETO
Lucas, Paul Derrick	PCO
Smith, Stephen John	CPOS
Lewis, Thomas Gregory	CPOD
Spencer, Robert George	POD
Cantlie, Ian Michael	SG1A
Crabb, Gary	SG1A
Hocking, Douglas Leonard	SG1A
McLennan, William	SG1A
Williams, Emlyn Gordon	ERPO
Ashfield, Mark James	Head Chef
Whalen, Amy Kerry	Chef
Osborn, Jeffrey Alan	Steward
Volosnuhina, Rita	Assistant Steward

Track Chart



Overview and Objectives – DY026a

Richard Sanders

DY26a formed part of the NERC Shelf sea Biogeochemistry programme. It had four principal objectives:

1. To continue the seasonal time series sampling at the three key sites (Shelf break, CaNDyFLoSS and Celtic Deep).
2. To provide sampling opportunities for the Shelf sea biogeochemistry students
3. To obtain samples of sinking particulate organic matter at differing stages of the tidal cycle to examine the role of tidal resuspension on elemental cycling beneath the thermocline.
4. To trial autonomous nitrate sensors CTDs and gliders as part of the Sensors on Gliders programme

Objective 1

We conducted CTD casts at each key site, sampling for DIC and nitrate concentrations to evaluate the differential uptake and re-mineralisation of these tracers, which is key to the operation of the shelf sea pump. In addition, we made at each site many of the rate measurements the programme needs to make to understand the differential elemental cycling.

Objective 2

Each SSB student (Matthew Bone, sediment coring, Richie Sims, near surface ocean profiler, Isabel Seguro, O₂/ Ar ratios, Kieran Curran, phytoplankton processes) got a good range of sampling opportunities at various sites in varying weather conditions).

Objective 3

Following the standard SSB observations outlined in objective 1 we undertook a highly temporally resolved timeseries of observations at the Celtic Deep. This comprised hourly CTDs with very high-resolution sampling near the bed coupled to hourly Snow Catcher deployments and near bottom respiration, bacterial production and nitrification measurements. Following this, we revisited the same site over a tidal cycle and collected near bed suspended material for similar biological rate measurements.

Objective 4

The Sensors on Gliders programme has integrated a nitrate sensor into a glider. This combination was trialed at the shelf break and on one further occasion during the cruise. Both deployments produced useful data. In addition, the nitrate sensor itself was deployed on the CTD in standalone mode on two occasions with extended bottle stops to allow reliable measurements to be made.

Narrative

Richard Sanders

3rd August 2014

Discovery slipped her moorings at 14:00 having completed fuel maintenance procedures. Engine propulsion trials carried out in the English Channel prior to dropping off an Engineer via boat transfer at Weymouth at approx 22:30.

4th August 2014

Discovery remained on passage, stopping just south of the Lizard to undertake trials of the PML Near Surface Ocean Profiler, large marine snow catcher and nets.

5th August 2014

Discovery arrived on station at the CaNDyFloSS site early in the morning and undertook a long day of station work similar to that which the main CaNDyFloSS programme will aim to undertake at three key sites, CaNDyFloSS itself, the shelf break and the Celtic Deep. This involved snowcatcher deployments throughout the water column, nets, productivity work, SAPS and coring.

6th August 2014

Discovery moved off the shelf break station into deep water in order to trial the combination of gliderbased nitrate sensor and new glider body-shape, which the Sensors on Gliders programme has been working on. This was followed by station work and a deployment of the PML Near Surface Ocean prior to glider recovery. Following the recovery *Discovery* undertook overnight winch trials.

7th August 2014

Conducted a long process station at the shelf break commencing 05:15 and concluding 23:55. This involved corers, nets, PML Near Surface Ocean Profiler, SAPS, snow catchers and CTDs.

8th August 2014

Undertook a long transit to the Celtic Deep.

9th August 2014

Undertook a long process station at the Celtic Deep similar to those conducted on 5th & 7th August.

10th August 2014

Having completed the basic suite of CaNDyFloSS sampling at each station, we undertook a 12h station with very highly resolved temporal sampling using the CTD and snow catchers both deployed every hour.

11th August 2014

The intense sampling on the previous two days resulted in a low activity day with just a 12:00 CTD, PML buoy deployment and nets & snow-catchers deployment.

12th August 2014

This day was devoted to obtaining samples of deep near bed particulate material in the Celtic Deep for biological rate measurements including respiration, bacterial production and organic phosphorus utilisation. We also deployed the PML Near Surface Ocean Profiler, which flooded on deployment and undertook a detailed study of particle fluxes out of the deep chlorophyll maximum.

13th August 2014

Undertook the regular noon CTD and obtained sediment cores for Matthew Bone. The PML Near Surface Profiler was deployed but failed to function satisfactorily. Following the 12:00 CTD we sailed for home.

14th August 2014

On passage. *Discovery* docked at National Oceanography Centre, Southampton 19:30.

Event Log

Stn	Gear	Site	Description	Date	Start time	Latitude	Longitude
001	CTD	Test south of Lizard	Test South Of lizard	04/08/2014	11:21	49 48.8	5 28.66
002	Buoy	Test south of Lizard	Test South Of lizard	04/08/2014	13:07	49 48.8	5 28.66
003	Buoy	Test south of Lizard	Test South Of lizard	04/08/2014	14:15	49 48.78	5 28.66
004	LMSC	Test south of Lizard	Test South Of lizard	04/08/2014	15:27	49 48.71	5 28.69
005	Net	Test south of Lizard	Test South Of lizard	04/08/2014	17:20	49 48.71	5 28.79
006	CTD	Candyfloss	Process Station	05/08/2014	05:10	49 23.18	8 37.13
007	LMSC	Candyfloss	Process Station	05/08/2014	06:05	49 23.18	8 37.13
008	LMSC	Candyfloss	Process Station	05/08/2014	07:25	49 23.18	8 37.13
009	SAPS	Candyfloss	Process Station	05/08/2014	08:00	49 23.18	8 37.13
010	SAPS	Candyfloss	Process Station	05/08/2014	09:45	49 23.10	8 37.13
011	CTD	Candyfloss	Process Station	05/08/2014	11:03	49 23.0	8 36.7
012	LMSC	Candyfloss	Process Station	05/08/2014	12:04	49 23.0	8 36.6
013	Net	Candyfloss	Process Station	05/08/2014	13:10	49 22.9	8 36.6
014	Net	Candyfloss	Process Station	05/08/2014	13:41	49 22.7	8 36.4
015	Net	Candyfloss	Process Station	05/08/2014	14:12	49 22.6	8 36.4
016	Net	Candyfloss	Process Station	05/08/2014	14:27	49 22.6	8 36.4
017	Net	Candyfloss	Process Station	05/08/2014	14:42	49 22.7	8 36.4

018	Net	Candyfloss	Process Station	05/08/2014	15:01	49 22.7	8 36.4
019	Net	Candyfloss	Process Station	05/08/2014	15:11	49 22.7	8 36.4
020	Net	Candyfloss	Process Station	05/08/2014	15:20	49 22.7	8 36.4
021	CTD	Candyfloss	Process Station	05/08/2014	16:10	49 22.3	8 36.48
022	Corer	Candyfloss	Process Station	05/08/2014	19:31	49 22.33	8 36.48
023	Corer	Candyfloss	Process Station	05/08/2014	19:57	49 22.33	8 36.48
024	Net	Candyfloss	Process Station	05/08/2014	21:10	49 22.33	8 36.44
025	Net	Candyfloss	Process Station	05/08/2014	21:53	49 22.33	8 36.44
026	Net	Candyfloss	Process Station	05/08/2014	22:02	49 22.33	8 36.44
027	Net	Candyfloss	Process Station	05/08/2014	22:21	49 22.33	8 36.39
028	Net	Candyfloss	Process Station	05/08/2014	22:28	49 22.33	8 36.12
029	Glider	Deep Glider Station	Deep Glider Station	06/08/2014	07:57	48 20.46	9 43.16
030	CTD	Deep Glider Station	Deep Glider Station	06/08/2014	08:23	48 20.3	9 43.63
031	SMSC	Deep Glider Station	Deep Glider Station	06/08/2014	12:58	48 20.31	9 43.63
032	Buoy	Deep Glider Station	Deep Glider Station	06/08/2014	13:30	48 20.32	9 43.62
033	LMSC	Shelf Break	Shelf Break	07/08/2014	05:15	48 34.2	9 30.6
034	LMSC	Shelf Break	Shelf Break	07/08/2014	05:48	48 34.2	9 30.6
035	CTD	Shelf Break	Shelf Break	07/08/2014	06:04	48 34.2	9 30.6
036	Net	Shelf Break	Shelf Break	07/08/2014	06:56	48 34.22	9 30.63

037	Net	Shelf Break	Shelf Break	07/08/2014	07:23	48 34.3	9 30.74
038	Net	Shelf Break	Shelf Break	07/08/2014	07:45	48 34.38	9 30.84
039	Net	Shelf Break	Shelf Break	07/08/2014	08:05	48 34.56	9 31.0
040	LMSC	Shelf Break	Shelf Break	07/08/2014	08:48	48 34.57	9 31.0
041	Corer	Shelf Break	Shelf Break	07/08/2014	09:25	48 34.57	9 30.97
042	Corer	Shelf Break	Shelf Break	07/08/2014	09:48	48 34.57	9 30.97
043	Corer	Shelf Break	Shelf Break	07/08/2014	10:10	48 34.57	9 30.97
044	CTD	Shelf Break	Shelf Break	07/08/2014	11:10	48 34.58	9 30.97
045	LMSC	Shelf Break	Shelf Break	07/08/2014	12:25	48 34.57	9 30.9
046	SAPS	Shelf Break	Shelf Break	07/08/2014	13:00	48 34.57	9 30.87
047	CTD	Shelf Break	Shelf Break	07/08/2014	15:00	48 34.3	9 30.3
048	Buoy	Shelf Break	Shelf Break	07/08/2014	16:12	48 34.27	9 30.27
049	Buoy	Shelf Break	Shelf Break	07/08/2014	18:54	48 34.16	9 33.33
050	Net	Shelf Break	Shelf Break	07/08/2014	20:40	48 34.66	9 35.17
051	Net	Shelf Break	Shelf Break	07/08/2014	21:09	48 34.85	9 35.48
052	Net	Shelf Break	Shelf Break	07/08/2014	21:24	48 34.9	9 35.52
053	Net	Shelf Break	Shelf Break	07/08/2014	21:41	48 35.15	9 35.47
054	Corer	Shelf Break	Shelf Break	07/08/2014	22:12	48 35.45	9 35.44
055	Corer	Shelf Break	Shelf Break	07/08/2014	22:37	48 35.38	9 35.26
056	CTD	Noon Station	Noon Station	08/08/2014	11:05	50 15.48	7 44.62
057	SMSC	Noon Station	Noon Station	08/08/2014	12:02	50 15.48	7 44.62

058	Buoy	Noon Station	Noon Station	08/08/2014	12:58	50 15.48	7 44.62
059	LMSC	Celtic Deep	Celtic Deep	09/08/2014	05:06	51 08.26	6 35.16
060	LMSC	Celtic Deep	Celtic Deep	09/08/2014	05:30	51 08.26	6 35.16
061	CTD	Celtic Deep	Celtic Deep	09/08/2014	06:08	51 08.26	6 35.16
062	LMSC	Celtic Deep	Celtic Deep	09/08/2014	06:42	51 08.26	6 35.16
063	SMSC	Celtic Deep	Celtic Deep	09/08/2014	07:06	52 08.26	6 35.16
064	Net	Celtic Deep	Celtic Deep	09/08/2014	07:32	53 08.26	6 35.37
065	Net	Celtic Deep	Celtic Deep	09/08/2014	07:56	51 08.20	6 35.59
066	Net	Celtic Deep	Celtic Deep	09/08/2014	08:13	51 08.09	6 35.91
067	Net	Celtic Deep	Celtic Deep	09/08/2014	08:28	51 08.00	6 36.07
068	Net	Celtic Deep	Celtic Deep	09/08/2014	08:39	51 07.93	6 36.11
069	SAPS	Celtic Deep	Celtic Deep	09/08/2014	09:05	51 07.84	6 36.33
070	LMSC	Celtic Deep	Celtic Deep	09/08/2014	10:22	51 07.29	6 37.22
071	CTD	Celtic Deep	Celtic Deep	09/08/2014	10:40	51 07.24	6 37.3
072	SMSC	Celtic Deep	Celtic Deep	09/08/2014	11:42	51 07.10	6 37.51
073	Corer	Celtic Deep	Celtic Deep	09/08/2014	12:00	51 07.10	6 37.52
074	Corer	Celtic Deep	Celtic Deep	09/08/2014	12:22	51 07.10	6 37.52
075	Corer	Celtic Deep	Celtic Deep	09/08/2014	12:40	51 07.10	6 37.52
076	CTD	Celtic Deep	Celtic Deep	09/08/2014	13:30	51 07.10	6 37.52
077	LMSC	Celtic Deep	Celtic Deep	09/08/2014	15:46	51 07.10	6 37.52
078	CTD	Celtic Deep	Celtic Deep	09/08/2014	16:04	51 07.10	6 37.52

079	Buoy	Celtic Deep	Celtic Deep	09/08/2014	17:37	51 07:09	6 37.51
080	Net	Celtic Deep	Celtic Deep	09/08/2014	17:37	51 08.94	6 38.49
081	Net	Celtic Deep	Celtic Deep	09/08/2014	20:38	51 08.86	6 38.56
082	Net	Celtic Deep	Celtic Deep	09/08/2014	20:45	51 08.64	6 38.73
083	Net	Celtic Deep	Celtic Deep	09/08/2014	21:04	51 08.61	6 38.77
084	Net	Celtic Deep	Celtic Deep	09/08/2014	21:13	51 08.53	6 38.87
085	Net	Celtic Deep	Celtic Deep	09/08/2014	21:22	51 08.44	6 38.98
086	CTD	Celtic Deep	Celtic Deep	10/08/2014	06:58	51 09.42	6 34.28
087	SMSC	Celtic Deep	Celtic Deep	10/08/2014	07:37	51 09.42	6 34.27
088	CTD	Celtic Deep	Celtic Deep	10/08/2014	08:00	51 09.38	6 34.52
089	SMSC	Celtic Deep	Celtic Deep	10/08/2014	08:39	51 09.38	6 34.52
090	CTD	Celtic Deep	Celtic Deep	10/08/2014	08:59	51 09.34	6 34.65
091	Corer	Celtic Deep	Celtic Deep	10/08/2014	09:31	51 09.23	6 35.04
092	CTD	Celtic Deep	Celtic Deep	10/08/2014	10:00	51 09.17	6 35.38
093	SMSC	Celtic Deep	Celtic Deep	10/08/2014	10:33	51 09.04	6 35.74
094	CTD	Celtic Deep	Celtic Deep	10/08/2014	11:03	51 08.97	6 35.91
095	SMSC	Celtic Deep	Celtic Deep	10/08/2014	11:48	51 08.86	6 36.2
096	CTD	Celtic Deep	Celtic Deep	10/08/2014	12:04	51 08.86	6 36.2
097	SMSC	Celtic Deep	Celtic Deep	10/08/2014	12:44	51 08.8	6 36.32
098	CTD	Celtic Deep	Celtic Deep	10/08/2014	12:59	51 08.8	6 36.32
099	SMSC	Celtic Deep	Celtic Deep	10/08/2014	13:42	51 08.75	6 36.4

100	CTD	Celtic Deep	Celtic Deep	10/08/2014	13:59	51 08.75	6 36.4
101	SMSC	Celtic Deep	Celtic Deep	10/08/2014	14:48	51 08.75	6 36.41
102	CTD	Celtic Deep	Celtic Deep	10/08/2014	15:08	51 08.74	6 36.4
103	SMSC	Celtic Deep	Celtic Deep	10/08/2014	15:30	51 08.74	6 36.42
104	CTD	Celtic Deep	Celtic Deep	10/08/2014	16:10	51 08.74	6 36.32
105	SMSC	Celtic Deep	Celtic Deep	10/08/2014	16:42	51 08.74	6 36.25
106	CTD	Celtic Deep	Celtic Deep	10/08/2014	17:00	51 08.74	6 36.25
107	SMSC	Celtic Deep	Celtic Deep	10/08/2014	17:42	51 08.74	6 36.25
108	CTD	Celtic Deep	Celtic Deep	10/08/2014	17:57	51 08.74	6 36.25
109	SMSC	Celtic Deep	Celtic Deep	10/08/2014	18:50	51 08.74	6 36.25
110	CTD	Celtic Deep	Celtic Deep	10/08/2014	19:06	51 08.74	6 36.25
111	Glider	Benthic A	Benthic A	11/08/2014	06:42	51 12.57	6 08.52
112	Buoy	Benthic A	Benthic A	11/08/2014	07:49	51 12.70	6 08.5
113	CTD	Benthic A	Benthic A	11/08/2014	10:58	51 12.70	6 08.5
114	SMSC	Benthic A	Benthic A	11/08/2014	11:47	51 12.70	6 08.5
115	SMSC	Benthic A	Benthic A	11/08/2014	11:59	51 12.70	6 08.5
116	Net	Benthic A	Benthic A	11/08/2014	12:31	51 12.70	6 08.5
117	CTD	Benthic A	Benthic A	11/08/2014	14:01	51 12.33	6 08.51
118	SMSC	Benthic A	Benthic A	11/08/2014	15:08	51 12.33	6 08.5
119	SMSC	Benthic A	Benthic A	11/08/2014	15:25	51 12.28	6 08.5
120	Net	Benthic A	Benthic A	11/08/2014	15:36	51 12.24	6 08.52

121	Net	Benthic A	Benthic A	11/08/2014	15:54	51 12.18	6 08.45
122	Net	Benthic A	Benthic A	11/08/2014	16:14	51 12.12	6 08.25
123	Net	Benthic A	Benthic A	11/08/2014	16:26	51 12.09	6 08.07
124	Net	Benthic A	Benthic A	11/08/2014	16:44	51 12.1	6 07.9
125	Net	Benthic A	Benthic A	11/08/2014	20:37	51 11.92	6 05.67
126	Net	Benthic A	Benthic A	11/08/2014	20:52	51 11.96	6 05.97
128	Net	Benthic A	Benthic A	11/08/2014	21:04	51 11.98	6 06.23
129	Net	Benthic A	Benthic A	11/08/2014	21:23	51 12.00	6 06.63
130	Corer	Benthic A	Benthic A	11/08/2014	21:51	51 12.02	6 07.04
131	Corer	Benthic A	Benthic A	11/08/2014	22:10	51 12.03	6 07.12
132	Corer	Benthic A	Benthic A	11/08/2014	22:36	51 12.02	6 07.21
133	SMSC	Celtic Deep	Celtic Deep	12/08/2014	05:00	51 08.91	6 36.25
134	SMSC	Celtic Deep	Celtic Deep	12/08/2014	05:24	51 08.91	6 36.25
135	CTD	Celtic Deep	Celtic Deep	12/08/2014	07:00	51 08.91	6 36.25
136	SMSC	Celtic Deep	Celtic Deep	12/08/2014	07:42	51 08.91	6 36.25
137	SMSC	Celtic Deep	Celtic Deep	12/08/2014	08:02	51 08.91	6 36.25
138	Net	Celtic Deep	Celtic Deep	12/08/2014	08:41	51 08.91	6 36.32
139	Net	Celtic Deep	Celtic Deep	12/08/2014	09:02	51 08.83	6 36.45
140	Net	Celtic Deep	Celtic Deep	12/08/2014	09:20	51 08.77	6 36.68
141	Net	Celtic Deep	Celtic Deep	12/08/2014	09:40	51 08.59	6 36.86
142	CTD	Celtic Deep	Celtic Deep	12/08/2014	11:00	51 08.39	6 37.32

143	SMSC	Celtic Deep	Celtic Deep	12/08/2014	11:50	51 08.33	6 37.58
144	SMSC	Celtic Deep	Celtic Deep	12/08/2014	12:05	51 08.3	6 37.62
145	SMSC	Celtic Deep	Celtic Deep	12/08/2014	12:18	51 08.25	6 37.7
146	CTD	Celtic Deep	Celtic Deep	12/08/2014	13:26	51 08.22	6 37.77
147	SMSC	Celtic Deep	Celtic Deep	12/08/2014	14:40	51 08.1	6 38.01
148	SMSC	Celtic Deep	Celtic Deep	12/08/2014	14:54	51 08.1	6 38.01
149	Buoy	Celtic Deep	Celtic Deep	12/08/2014	15:27	51 08.10	6 38.00
150	CTD	Celtic Deep	Celtic Deep	12/08/2014	18:00	51 08.10	6 38.00
151	SMSC	Celtic Deep	Celtic Deep	12/08/2014	18:40	51 08.10	6 37.9
152	SMSC	Celtic Deep	Celtic Deep	12/08/2014	19:05	51 08.10	6 37.9
153	Net	Celtic Deep	Celtic Deep	12/08/2014	20:35	51 08.10	6 37.96
154	Net	Celtic Deep	Celtic Deep	12/08/2014	20:52	51 07.97	6 38.11
155	Net	Celtic Deep	Celtic Deep	12/08/2014	21:09	51 07.95	6 38.24
156	Net	Celtic Deep	Celtic Deep	12/08/2014	21:24	51 07.88	6 38.29
157	Corer	Benthic A	Benthic A	13/08/2014	06:37	51 12.28	6 07.3
158	PML Buoy	Benthic A	Benthic A	13/08/2014	07:18	51 12.28	6 07.3
159	CTD	Benthic A	Benthic A	13/08/2014	11:00	51 12.57	6 03.36

Sensors on Gliders

Steve Woodward (NOCS MARS Group), Sam Ward (NOCS MARS Group) and John Walk (NOCS OTE Group)

Objectives

For the Sensors on Gliders project, DY026 was a technology-proving cruise to practice deploying the NOC's Lab-On-Chip (LOC) nitrate sensors on a glider at sea.

The primary objectives were:

- to prove that the glider could be operated successfully with the 3.5Kg sensor-pair payload
- to prove that the base station, glider and sensor could communicate successfully at sea
- to develop optimal sampling patterns for operating the sensors on the glider

A secondary objective was to deploy the sensor on the CTD frame at every opportunity to gather more field data for this relatively new technology.

There were no science objectives for the Sensors on Gliders project on this cruise.

Equipment

2 x NOC LOC Nitrate sensors in a single housing with external oil bladder and no battery

1 x NOC LOC Nitrate sensor housed with internal battery and oil bladder, and a CTD bottle clamp

1 x NOC LOC Nitrate sensor (spare)

2 x Kongsberg Seagliders (SG534 + SG533 spare)

Sensor

The NOCS Lab-On-Chip nitrate sensor is one of a suite for sensors developed by the OTE Group at NOCS for different chemistries using microfluidic technology. The nitrate sensor allows in-situ measurement of nitrate+nitrite (or nitrite only) with a limit of detection of $0.025\mu\text{M}$ (nitrate) and $0.02\mu\text{M}$ (nitrite) and uses very small quantities of reagent (Beaton, 2012).



Inputs of sea water sample, artificial sea water blank or $10\mu\text{M}$ potassium nitrate standard are sequentially combined with Imidazole buffer and passed through a cadmium column to convert nitrate to nitrite, then combined with Griess reagent to develop a colour which is measured by absorption of light from a 525nm LED. The results from the sample, standard and blank are combined to give the nitrate+nitrite result. The chemical processing is done in-situ, so for example when deployed on the CTD frame, the chemistry is complete and raw results are available when the frame is lifted from the water.

The mixing cells, reaction cells and measurement channels are all contained in a microfluidic chip, so called because the central layer superficially resembles an electronic printed circuit.

The picture shows a disassembled sensor with the chip at the base and the electronics, valves and syringe pumps fixed directly to it. The chemicals are stored externally in blood bags and connected to the opposite face of the chip. The external housing varies to suit the platform on which the sensor is deployed.

The total pumping and reaction time with the current technology is around 6.5 minutes for each input giving about 20 minutes for a blank-sample-standard pattern. In some contexts, for example long-term monitoring of a river, this is not a problem, but holding up a CTD cast with long stops at depth is inconvenient, and holding the Seagliders at depth (loitering) is tricky. The sensor is programmable, so we tried five different sampling patterns on this cruise, all designed to reduce the sampling interval. They were as follows:

PATTERN 1	PATTERN 2	PATTERN 3	PATTERN 4	PATTERN 5
wait until in water	wait until in water	start immediately	start immediately	start immediately
BLANK	BLANK	BLANK	BLANK	BLANK
SAMPLE	SAMPLE	STANDARD	STANDARD	STANDARD
STANDARD	STANDARD	BLANK	BLANK	BLANK
repeat whole pattern for rest of cast	SAMPLE	STANDARD	STANDARD	STANDARD
	repeat whole pattern for rest cast	wait until below surface	wait until depth exceeds 10m	wait until depth exceeds 10m
		SAMPLE	STANDARD	SAMPLE
		repeat sample to end of dive	repeat sample to end of dive	SAMPLE
		BLANK	BLANK	BLANK
		STANDARD	STANDARD	repeat SSB to end of deployment
		BLANK	BLANK	
		STANDARD	STANDARD	
		wait for ascent	wait for ascent	
		SAMPLE	SAMPLE	
		repeat sample to just below surface	repeat sample to just below surface	
		BLANK	BLANK	
		STANDARD	STANDARD	
		BLANK	BLANK	
		STANDARD	STANDARD	

PATTERN 1 gives the best nitrate results but each iteration takes 3x6.4 minutes which means on a CTD cast (where the sensor is running continuously) you need to hold the CTD at each required depth for 4x6.4 minutes to guarantee a complete set of blank-sample-standard. The wait at the start is to avoid drawing in air on the deck. We used this pattern in Deployment 2.

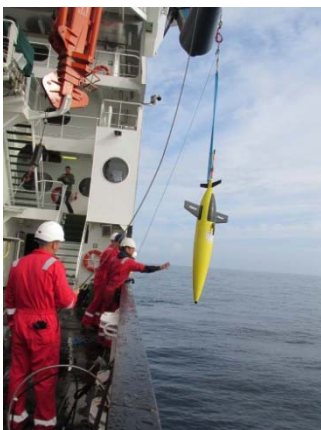
PATTERN 2 is an attempt to reduce the time between obtaining samples to try to get results from a CTD cast moving at normal speed. Each sample has a neighbouring blank but is not bracketed by blank+standard. We used this in Deployment 3 (with no stops) and Deployment 4 (with 10 minute stops).

PATTERNS 3 & 4 designed to run as a pair on two sensors on the Seaglider. One sensor is doing samples for the whole deployment except for a blank+standard at the start and end of the descent and ascent. The other sensor is doing standards the whole time. This pattern requires enough time at the top and bottom of the dive to complete the bracketing blank+standard and at 0.1m/s dive speed that means at least 42m at the top and bottom, so it is only practical if there is sufficient depth of water. It also fails if the Seaglider does not reach its target depth hence this pattern was only used on Deployment 1 in 1500m of water.

PATTERN 5 was designed on the cruise to solve two problems. Firstly we were to remain at the shallow benthic sites (<100m) for the remainder of the cruise and secondly we only had one functioning sensor for the Seaglider. The results from Deployment 2 suggest that there is no depth-dependence on the results of standards measurements with this sensor, so it needs to be done at the start of deployment only, allowing us to reduce the time between sampling. We used this pattern in Deployment 5 and would have tried it (without the depth check) on a further CTD deployment but unfortunately the sensor failed before we were able to try it.

Glider

The Kongsberg Seaglider is an autonomous underwater vehicle that has no direct propulsion but instead rely on the forward motion generated by small wings as they descend or ascend in the water. This saving in power allows them to be deployed for extended periods.



They control their buoyancy by pumping oil in and out of an external bladder and they pitch and roll by moving their battery around to alter their trim. They are capable of navigating from one waypoint to another in a series of dive profiles, and they return data and gather new instructions each time they surface by communicating via Iridium to a base station. These instructions can set new mission parameters and sensor settings as well altering the trim of the Seaglider based on the telemetry from the previous dive. Piloting the Seaglider (sending these instructions) was done from NOC, as a reliable Internet connection is needed to upload the command files.

The Seaglider has a large free-flooding payload bay, which is sufficient to house the sensor on a custom mounting. Connection to the Seaglider is via IE55 serial cables, which carry power and RS232 serial communications. For this cruise, we configured the Seaglider to send its GPS time (to allow us to correlate

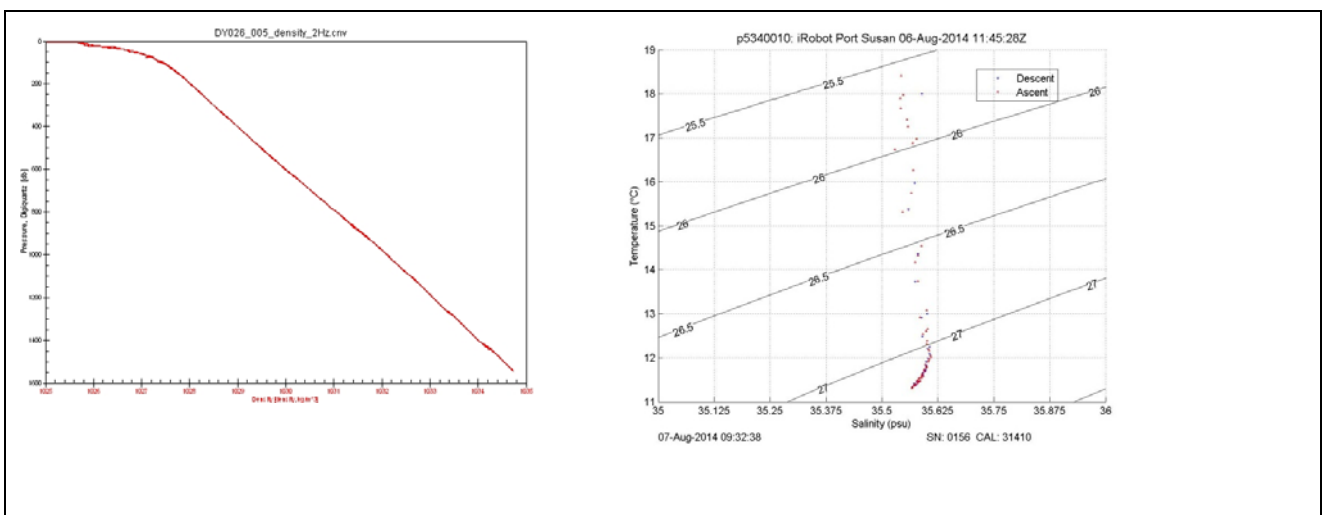
the sensor data with the Seaglider's own sensor data) at the start of the deployment and depth+direction (i.e. ascent and descent) every five seconds to allow the sensor to make depth-dependent decisions in its processing (see PATTERNS 4 & 5 above). It is also possible to include three arbitrary sensor parameters in the command file sent to the Seaglider and we used those to set the cut-off depths used in the sampling logic by the sensor.

Installation of dual Lab-on-Chip (LOC) Nitrate sensors onto a Seaglider requires a more extensive re-ballasting procedure than is normal before a deployment due to both their size and weight (2548cm³ and 3635g for the combined sensor housing, plus bags and cables). Using an Ogive fairing with increased payload capacity in the aft fairing was essential. To compensate for the negative buoyancy of the sensor, TG-42 syntactic foam pieces were added (384.3g in strips around the circumference of the battery hull and 600g in machined blocks bolted to the aft fairing top hatch cover).

Aiming for a target density (ρ) of 1026.8kg/m³ at 1000m depth, ballasting was checked by weighing the Seaglider (SG534) – first dry, then suspended in a freshwater tank with its buoyancy device (VBD) pumped to maximum and minimum volume. Additionally, the Seaglider was deployed tethered from the marina at NOCS. Erring on the side of caution, 728g of brass nose weight and 150.5g of lead ballast added to give a calculated thrust of 50-100cm³ at ρ . Pitch, VBD and roll centres measured during the dock tests.

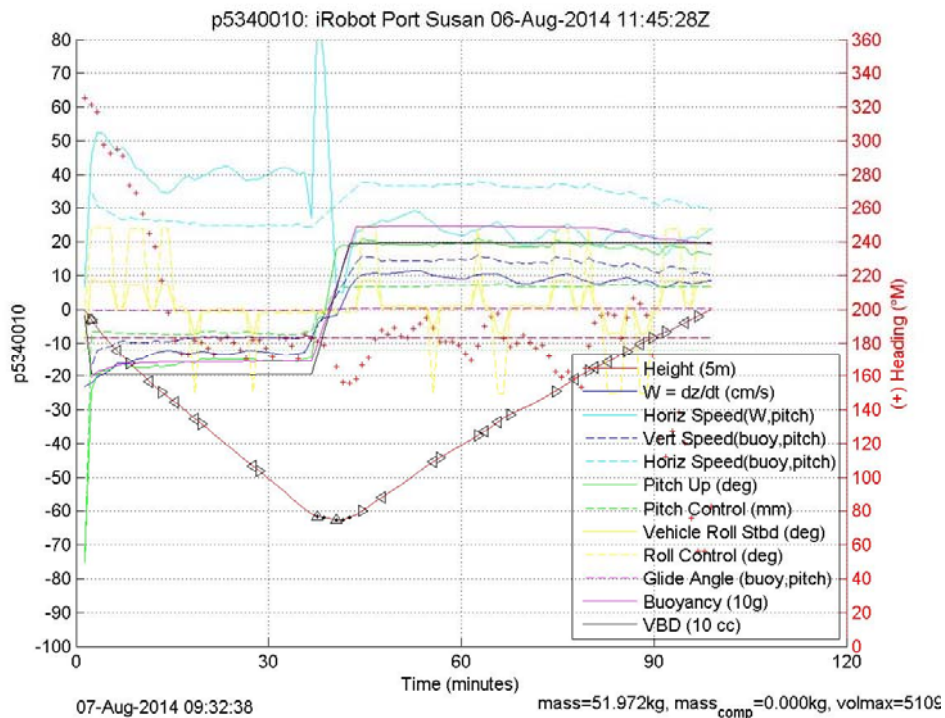
The density profile from CTD002 (Central Celtic Sea processing station, 04/08/2014) showed a surface density of 1025.3kg/m³, rising to 1027.6kg/m³ at 100m. In anticipation of the SG534 therefore being too buoyant and unable to dive to >100m, 136.2g of lead ballast was added to the aft fairing, giving a calculated 150cm³ of thrust at a new ρ of 1027.8kg/m³.

SG534 was not as buoyant as predicted, probably due to inaccuracy of the Seaglider CT sensor (SBE s/n 0156). Between dives 6 and 10, the centre point of the VBD (\$C_VBD) had to be adjusted from 3498 to 2798 A/D counts, a change of 170cm³.



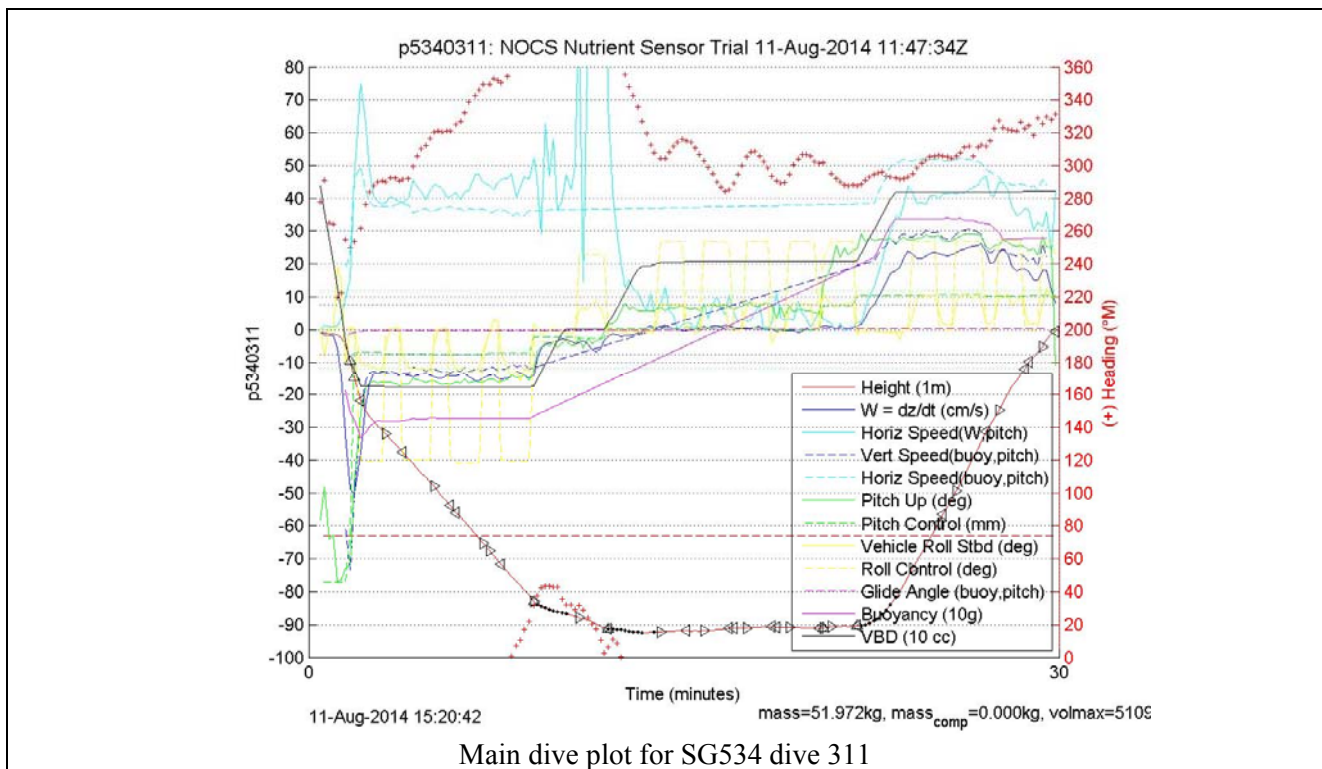
Deployed CTD005 and SG534 dive 10 density profiles. Seaglider density at apogee (~300m) = 1027.2kg/m³.
 Deployed CTD density at 300m = 1028.6kg/m³.

The main dive plot for dive 10 shows a roughly symmetrical profile. Vertical velocity is quite stable in both the dive phase (~12cm/s) and climb phase (~10cm/s). The apogee maneuver is reasonably smooth, with only a minor drop in horizontal speed. After the initial shallow dives, SG534 made steady progress towards its waypoint. Although some further adjustment is required, from this point there is no doubt that SG534 could be trimmed to perform well over a longer deployment in this area. It is therefore clear that the Ogive fairing Seaglider is a suitable platform for deployment of the LOC Nitrate sensor.



Main dive plot for SG534 dive 10

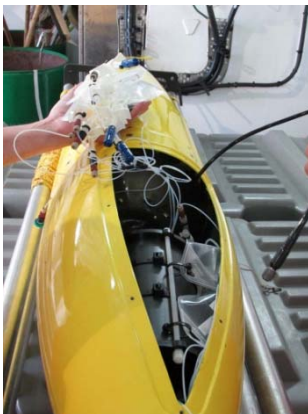
One function of the Seaglider platform, which could be potentially useful for LOC sensor sampling strategies, is the ability to loiter at depth. Using the \$T_LOITER parameter, SG534 was held at 90m for 10 minutes on dive 311.



Deployment 1

6 August (J14218) 48°20' N, 9°43' W in 1500m water: Seaglider test dives to 50m, 300m and 500m

This was the first Seaglider deployment with the sensor. The sensor was loaded into the payload bay the night before, about an hour's work to get the sensors and all the bags organized and tied down. The sample inlet tubs passed through the fairing with the filters on the outside.

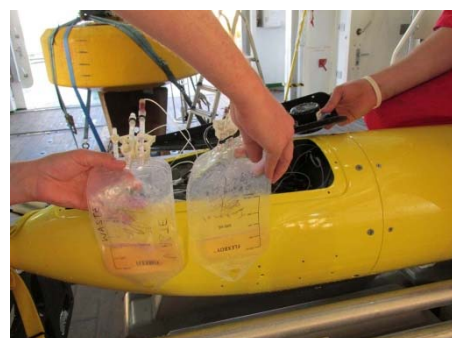


Pre-flight system checks carried out on deck and these took about 45 minutes on this deployment while the actual launch of the Seaglider took less than 10 minutes. The Seaglider remains with its communication antenna just out of the water until it receives its instruction to dive and we waited to see the Seaglider dive successfully before moving the ship back to the day's process station about 1 nautical mile way (to avoid any collisions).

The first test dives went well and the Seaglider flew better than expected with the heavy sensor installed.

Both sensors were enabled for the 300m and 500m dives, deployed as a pair using sampling PATTERNS 3 & 4 as described earlier.

Data for both dives was retrieved from the Seaglider when it was recovered, together with the summary data downloaded by the Seaglider itself from the sensors at the end of each dive. The communications with the sensor worked perfectly and both sensors operated correctly for the 300m dive and the first part of the 500m dive. The data has yet to be processed back at NOC. Sadly one of the sensors (the one running PATTERN 3)



failed with a pump jam at about 300m. We attempted a repair using parts from the spare sensor but it did not appear to be returning good results when tested on the ship with a standard as input and so was not deployed again (although it went back out as a passenger in Deployment 5 so as not to alter the buoyancy of the Seaglider).

An incorrect setting (RECORDABOVE) in the command file for the 300m dive meant that the Seaglider told the sensors to stop at 100m and ceased sending it status messages. The sensors ignore stop messages from the Seaglider so in fact they have data all the way down to 300m but the depth is not known and the bottom blank-standard sequence did not run, highlighting a significant flaw in PATTERN 3 which is addressed by PATTERN 5.

The 500m dive was aborted by the Seaglider shortly after beginning its ascent. Therefore this dive yielded data in the first part of the dive only. The Seaglider was running beta software so considered imprudent to deploy it again without an investigation by the manufacturer into what went wrong and whether or not it was safe to redeploy the Seaglider. Happily, we did in Deployment 5.

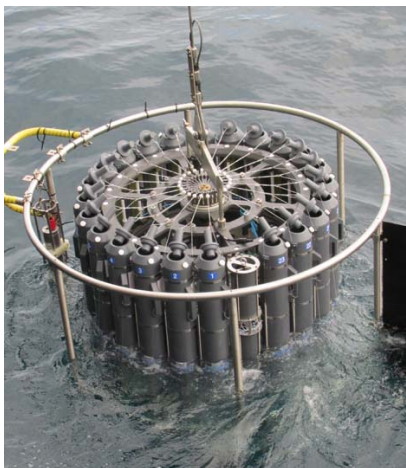
Deployment 1						
SG Dive #	Event	Longitude N	Latitude W	Date	Time UTC	Depth
	Deployed	4820.440	0943.190	060814	0658	
6	Dive Start	4820.407	0943.224	060814	0805	85
	Dive End	4820.421	0943.224	060814	0829	
7	Dive Start	4820.367	0943.041	060814	0841	90
	Dive End	4820.392	0943.123	060814	0906	
8	Dive Start	4820.354	0943.181	060814	0917	100
	Dive End	4820.263	0943.287	060814	0958	
9	Dive Start	4820.246	0943.352	060814	1004	225
	Dive End	4820.196	943.729	060814	1133	
10	Dive Start	4820.254	943.856	060814	1145	190
	Dive End	4819.596	943.674	060814	1345	
11	Dive Start	4819.742	-943.660	060814	1356	500
	Dive End	4819.246	-943.643	060814	1747	
	Recovery	4818.770	-942.790	060814	1830	

SG534 deployed from the forward auxiliary winch on the P frame using the rigid rope deployment rig. Conditions were calm and a buoyancy test was undertaken. Once the deployment team were satisfied that SG534 was sitting in the correct position whilst in the water it was deployed and began its mission. The

recovery went very smoothly due to the calm conditions and SG534 was recovered using the mid ships crane once the rudder was noosed using the carbon fiber recovery pole and aluminum hoop.

Deployment 2

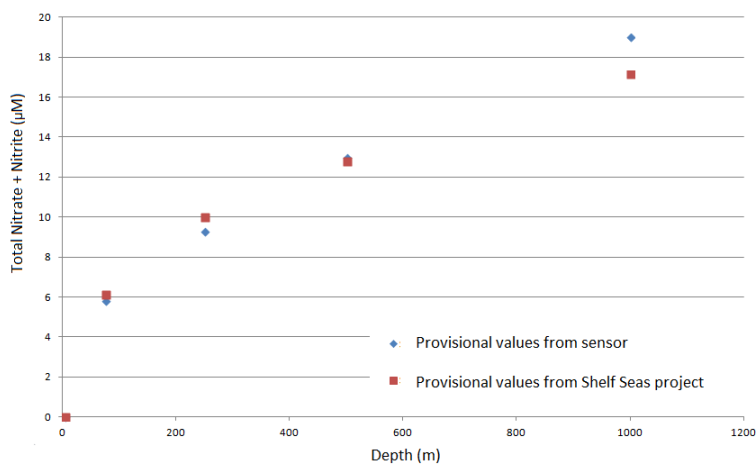
6 August (J14218) 48°20'N, 9°43' W in 1500m water: CTD cast to 1500m (with 25 minute stops at 1000m, 500m, 250m, 75m and 5m but the sensor running continuously for the whole cast)



This was the first CTD deployment of the sensor on this cruise. One improvement to the sensor housing design over previous cruises was the addition of a bottle clamp allowing it to be easily swapped for a bottle on the CTD bottle carousel. This worked well although it was suggested a handle on the front (as the bottles have) might make it slightly easier. The top half of the sensor housing in the picture contains the blood bags and the bottom half contains the sensor, the pressure-compensating bladder and a battery.

With PATTERN 1 sampling, the sensor has to be started (by attaching a shorted IE55 terminator) shortly before it enters the water to ensure it doesn't draw in air. For this deployment we allowed half an hour; in subsequent deployments we allowed 10 minutes and in all cases the CTD was in the water within 2 minutes of starting the sensor, so the delay is unnecessary as the first blank cycle takes 6.4 minutes.

This was the most successful and informative deployment of the sensor on the cruise as it was our only opportunity to put it onto the CTD in deep water. The CTD cast went to 1500m and the sensor was logging continuously throughout. However, we also halted the CTD at five locations long enough to complete 1 cycle of the CTD FAST sampling pattern. The provisional results (shown below) correlate quite well with the provisional lab-based nitrate values produced for the Shelf Seas project on the same cruise using water from the same CTD cast.



Two other interesting features from these results are an indication that the nitrate values obtained for the standard are not depth-dependent so it may be sufficient to process the standard a couple of times at the start of the dive, increasing the possible sampling rate. In addition, nitrate values for the depths between the 25-

minute stops appear to correlate well with those where we stopped so sampling on the move looks feasible at least when nitrate values are changing relatively slowly and continuously with depth. (The sensor is only actually drawing in seawater for about 2.5 minutes in the 20 minute cycle which may explain why the results were reasonable).

The electronic components in the sensor are pressure rated to 6000m and the sensor returned to the surface fully functional. Shortly after the deployment one electronic component (Dallas Semiconductor DS1374 real time clock) did report a failure and we swapped out the processor board out for a spare to be able re-deploy the sensor a couple of days later. In subsequent testing however the board seemed to be functioning correctly so possibly the fault was due to a faulty connection rather than the collapse of a component.

Deployment 3

8 August (J14220) 50°15' N, 7°44'W in 106m water: CTD cast to 100m (with no stops)

This was a first attempt to deploy on a CTD without stopping, but in 100m water, only one complete sample achieved showing the limitations of sampling PATTERN 2 in shallow water. The result is yet to be processed. From the CTD results we would expect a surge in nitrate (to about 8 μ M) at around 45m and deeper.

Deployment 4

9 August (J14221) 51°7'N, 6°37'W in 103m water: CTD cast to 85m (with 10 minute stops at 85m,75m,65m,55m,45m,35m,25m,15m,10m,5m)

This deployment made to see if a compromise of using PATTERN 2 (with at most 6.5 minutes between samples) and stopping for 10 minutes at selected depths would produce good results. They should also provide a good reference for the glider operations in the next deployment. Bottle samples also processed for us by the Shelf Seas Biogeochemistry project giving provisional figures for the same depths.

We would have expected to see a significant drop in the nitrate levels at around 40m due to a spike in activity of phytoplankton during rough weather.

Unfortunately the results from the sensor indicate that something failed in the optical processing, either the LED failed or the optical cell became blocked, so no sensor results are available for this cast.

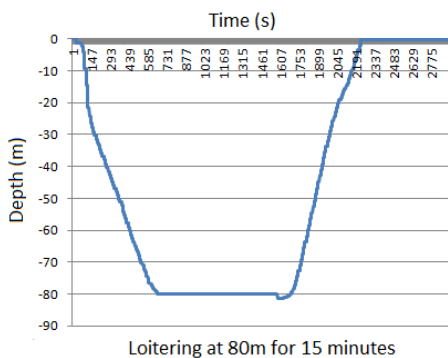
Deployment 5

11 August (J14223) 51°7'N, 6°37'W in 103m water: Seaglider “loitering” trials to 60m

This second deployment with the Seaglider aimed to test the Seaglider’s “loitering” capability. The Seaglider is theoretically capable of holding a particular depth for a period of time during its normal dive profile. This suits the nitrate sensor because we can process a full sample at a specific depth, just as we do when the CTD is halted. For this deployment we designed a new sampling pattern where the standards were run at the start of the dive profile only, then two samples to each blank for the remainder. We retained the depth check at the start to avoid sucking in surface bubbles (although this may not be necessary) but the other two depth

parameters were not used. Both sensors were deployed to avoid altering the Seaglider's buoyancy, but only one was enabled in the command file.

Communications with the piloting team at NOC were hampered by the Iridium phone getting a poor signal and several iterations of the pre-flight checks were required to establish that the Seaglider had the correct parameters. To be able to do this from the ship would make the task significantly easier, at least for trial deployments.



The Seaglider uses an altimeter to avoid colliding with the seabed and several dives were required to get this working properly (the altimeter gives false returns at a range of 5-8m until sensitivity is set correctly). However, one dive was successfully completed with the Seaglider loitering at depth for just over fifteen minutes, which is a good result (the plot is taken from the depths reported to the sensor by the glider).

In quite lively seas the Seaglider was located and recovered as quickly and smoothly as before. As with Deployment 1, all communications between Seaglider and sensor worked correctly and a full set of files retrieved from the Seaglider on completion of the dive. This showed the sensor had correctly executed the sampling pattern. Unfortunately the results from the chemistry are clearly not valid and an extended post-deployment test of the sensor has produced similar results. The cause is not identified.

Deployment 5						
SG Dive #	Event	Longitude N	Latitude W	Date	Time UTC	Depth
	Deployed	5112.679	608.471	11/08/14	0642	
304	Dive Start	5112.714	608.392	110814	0650	30
	Dive End	5113.269	608.093	110814	0727	
305	Dive Start	5113.439	607.966	110814	0739	30
	Dive End	5113.860	607.846	110814	0815	
306	Dive Start	5113.932	607.828	110814	0822	45
	Dive End	5114.182	607.929	110814	0854	
307	Dive Start	5114.215	607.926	110814	0900	70
	Dive End	5114.167	608.196	110814	0936	
308	Dive Start	5114.158	608.195	110814	0941	95
	Dive End	5114.007	608.764	110814	1013	
309	Dive Start	5113.957	608.763	110814	1021	70
	Dive End	5113.623	609.110	110814	1057	

310	Dive Start	5113.492	609.105	110814	1108	90
	Dive End	5113.318	609.421	110814	1138	
311	Dive Start	5113.180	609.399	110814	1147	90
	Dive End	5111.240	609.015	110814	1349	
312	Dive Start	5111.147	608.935	110814	1354	85
	Dive End	5110.811	608.330	110814	1445	
313	Dive Start	5110.662	608.150	110814	1455	85
	Dive End	5110.639	607.347	110814	1554	
314	Dive Start	5110.607	607.269	110814	1559	87
	Dive End	5110.975	606.509	110814	1658	
315	Dive Start	5110.997	606.443	110814	1703	82
	Dive End	5111.294	606.167	110814	1726	
	Recovery	5111.850	605.69	110814	1815	

The same methods for deployment and recovery of the Seaglider were used as in Deployment 1. Conditions were worse (sea state 4-5). The deployment team made the decision to proceed, having confidence in the ships station holding capabilities and its maneuverability in recovery operations.

SG534 was recovered in sea state 5 - 6, the ship was impressive in positioning itself next to the Seaglider in a large swell. In a slight variation from the first recovery, The recovery line was tied to an eye on the bulwark and a boss hook was attached which enabled the Seaglider to be pulled out of the water at double the speed.

Successes

We have demonstrated that it is possible to run the LOC nitrate sensors on a Seaglider at sea. The Seaglider operated well and the communications between the Seaglider and the sensor worked perfectly.

The data we have gathered from the two Seaglider deployments should be sufficient to design optimal sampling strategies for operating the sensors on the Seaglider in both deep and shallow waters.

We have demonstrated that it is now very easy to deploy the sensor onto the CTD frame and by picking a strategy that does not require long stops at each depth, we can operate on that platform whenever there is a bottle slot spare with no significant impact on CTD operations.

The deployment on the deep CTD cast (to 1500m) has given us valuable experience of operating the sensor at significant depth and the correlation of the nitrate values obtained by the sensor and those obtained from the water samples gives us further confidence in this sensor technology. It has also shown that standards measurements do not vary with depth with this sensor, so it should be sufficient to take a couple of standards measurements at the start of a CTD cast or Seaglider profile and then iterate samples and blanks, achieving a higher sampling rate than was thought possible.

Room for improvement

The discrepancy between the provisional nitrate values obtained on the deep CTD deployment and those obtained by conventional processes from the water samples will need further investigation.



The cause of the failure of the sensor's real time clock and syringe pump presumably due to pressure will also need further explanation.

The requirement to fill the sensor housing with oil (to balance the sea pressure) makes the job of fixing the sensors at sea much harder than it would otherwise be, because the housing needs careful re-sealing and testing before it can be re-deployed.

The sensor fits in the Seaglider's payload bay with room to spare, especially to the sides of it and in the tail. The most time-consuming aspect of the installation was securing the blood bags. If these can be contained in some way (as they are in the CTD sensor housing) the process may be speeded up.

Alternatively, if running with a single sensor is considered to be adequate, it might be possible to locate the bags behind the sensor in the tail space with no need to secure them.

Ways to reduce the processing time need to be explored further to reduce the sampling interval to operate the sensor more effectively on moving platforms like the CTD and the Seaglider. This will be achieved by a mixture of new technology (we are investigating new pump seals that should reduce flushing time) and alternative sampling strategies.

It should be possible to get sensor data back via the Seaglider and Iridium to the base station but this has yet to be proven. Whilst it would not be possible to recover all of the raw data from the sensor in this way, it certainly is possible to get a status report and some averaged results - on this cruise we demonstrated getting that data back as far as the Seaglider.

Acknowledgements

Thanks to Malcolm Woodward for providing the nitrate figures and processing an extra set of bottles for us in Deployment 3. Also to our colleagues in the MARS group back home for piloting the Seaglider at unsociable hours and our colleagues in the OTEG group for ongoing support and for processing the data. Finally, thanks to the technicians and scientists on the cruise for allowing us the opportunity to conduct these trials.

Nutrients

Malcolm Woodward, Plymouth Marine Laboratory, UK

Objectives

To investigate the spatial and temporal variations of the micromolar nutrient species; Nitrate, Nitrite, Silicate, Ammonium and Phosphate during the DY026 research voyage in the Celtic Sea and Western Approaches off the West coast of the UK. Carry out nutrient analysis from zooplankton and benthic experiments where required as part of the SSB programme (Giering and Bone).

Please see individual cruise reports for these colleagues as to their individual sampling protocols.

Sampling and Analytical Methodology

Sample preparation and procedure

There was minimal storage of the Underway non-toxic and CTD water column samples except for the time waiting to be analysed in the laboratory. These samples were always run at lab temperature and were not filtered. 60m ml HDPE Nalgene bottles were used for all the nutrient sampling, these were aged, acid washed and cleaned initially, and stored with a 10% acid solution between sampling. Samples were taken from the Sea-Bird CTD system on-board the RRS Discovery. The sample bottle was washed 3 times before taking final sample, and capping tightly. This was then taken immediately to the analyzer in the lab and analysis conducted as soon as possible after sampling. Nutrient free gloves (Duratouch) were used and other clean handling protocols were adopted as close to those according to the GO-SHIP protocols, (2010).

Sample Analysis

The micro-molar segmented flow auto-analyser used was the PML 5 channel (nitrate, nitrite, phosphate, silicate and ammonium) Bran and Luebbe AIII system, using classical proven analytical techniques.

The instrument was calibrated with home produced nutrient standards and then compared regularly against Nutrient Reference Materials, from KANSO Technos, Japan. The results from this also being part of a global nutrient programme (the INSS, International Nutrient Scale System) to improve nutrient analysis data quality world-wide.

The analytical chemical methodologies used were according to Brewer and Riley (1965) for nitrate, Grasshoff (1976) for nitrite, Kirkwood (1989) for phosphate and silicate, and Mantoura and Woodward (1983) for ammonium.

References

Brewer P.G. and Riley J.P., 1965. The automatic determination of nitrate in seawater. *Deep Sea Research*, 12, 765-72.

Grasshoff K., 1976. Methods of seawater analysis. *Verlag Chemie, Weinheim and New York*, 317pp.
 Kirkwood D., 1989. Simultaneous determination of selected nutrients in seawater. *ICES CM 1989/C:29*.
 Mantoura, R.F.C and Woodward E.M.S, 1983. *Estuarine, Coastal and Shelf Science*, 17, 219-224.

Water samples taken from the 24 x 10litre Stainless Steel CTD/Rosette system (SeaBird). Clean handling techniques employed to avoid any contamination of the samples, particularly for the ammonium samples. Gloves used were Dura-Touch to minimize nutrient contamination. Samples were kept tightly closed until just before analysis for the ammonium, this to avoid any contamination from external sources.

Table. CTD Samples Analysed by AAIII Micromolar Analysis

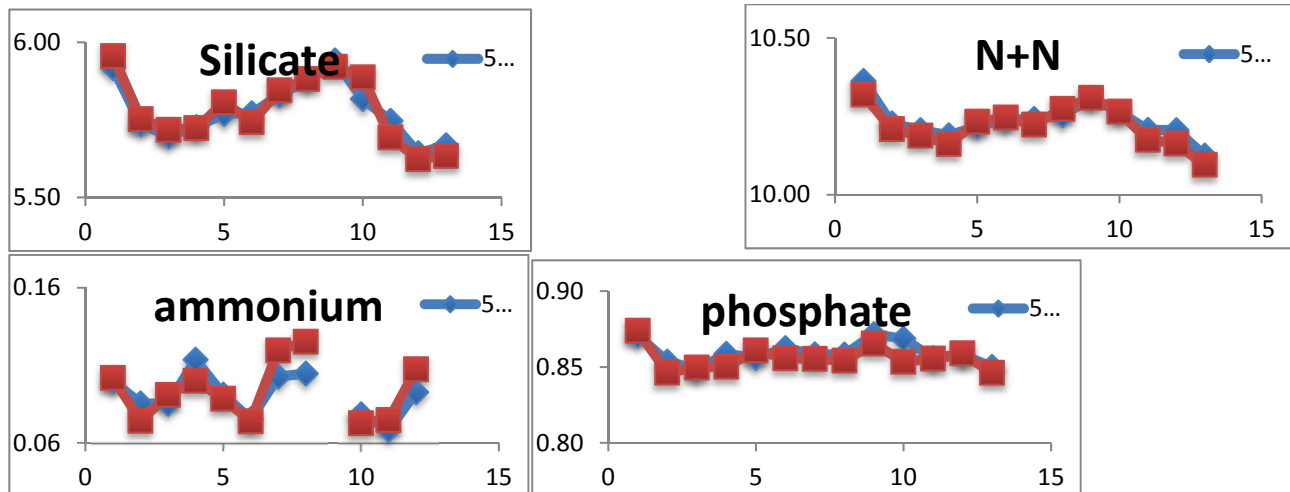
Date	CTD	Station	Position	CTD bottle analysed
04/08/14	CTD_001	001	49 ^o 48.80'N 5 ^o 28.66'W	Bottles 1,5,7,10,14,16,18,20 (depths: 80, 65, 50, 35, 25, 17, 17, 5m)
05/08/14	CTD_002	006	49 ^o 23.194'N 8 ^o 37.129'W	Bottles 4,7, 10, 13, 16, 19, 24 (depths: 136, 100, 60, 45, 30, 20, 5m)
05/08/14	CTD_003	011	49 ^o 23.012'N 8 ^o 36.70'W	Bottles 3, 5, 8, 12, 17, 21 (depths: 137, 100, 50, 34, 20, 5m)
05/08/14	CTD_004	021	49 ^o 22.311'N 8 ^o 36.465'W	Bottles 1,4,6,9,10, 16, 19, 22 (depths: 136, 100, 60, 45, 40, 30, 20, 5m)
06/08/14	CTD_005	030	48 ^o 20.30'N 9 ^o 43.62'W	Bottles 2,3, 4,5, 6, 7, 11, 16, 18, 19, 23 (depths: 1000, 500, 250, 125, 75, 50, 35, 26, 20, 10, 5m)
07/08/14	CTD_006	035	48 ^o 34.193'N 9 ^o 30.616'W	Bottles 1, 4, 7, 11, 15, 19, 22 (depths: 197, 120, 50, 40, 35, 15, 5m)
07/08/14	CTD_007	044	48 ^o 34.193'N 9 ^o 30.616'W	Bottles 1,3,6,9,11,15,17,21 (depths: 170, 120, 90, 55, 42, 30, 20, 5m)
07/08/14	CTD_008	047	48 ^o 34.27'N 9 ^o 30.27'W	Bottles 2,4,7, 10, 13, 16, 19, 24 (depths: 195, 120,90,60,40,29,15,5m)
08/08/14	CTD_009	056	50 ^o 15.48'N 7 ^o 44.61'W	Bottles 1,3,7,9,12,15,18,22 (depths: 93,60,45,36,34,32,20,5m)
09/08/14	CTD_010	061	51 ^o 08.26'N 6 ^o 35.14'W	Bottles 1,4,7,11,15,18,19,22 (depths: 85,60,50,36,30,20,10,5m)
09/08/14	CTD_011	071	51 ^o 07.748'N 6 ^o 37.33'W	Bottles 1,3,5,8,16,17,19,21,1 (depths: 92,70,50,36,34,28,15,5m)
09/08/14	CTD_012	076	51 ^o 07.099'N 6 ^o 37.499'W	Bottles 1,2,3,4,5,6,7,8,9,10(depths: 85,75, 65,55,45,35,25,15,10,5m)
09/08/14	CTD_013	078	51 ^o 07.099'N 6 ^o 37.499'W	Bottles 14,8,12,14,16,19,22(depths: 92,65,48,39,37,25,15,5m)

10/08/14	CTD_014	086	51°09.42'N 6°34.27'W	Bottles 1,4,5,6,7,8(depths: 92,87,82,77,30.5,5m)
10/08/14	CTD_015	088	51°09.415'N 6°34.252'W	Bottles 9,2,13,14,16,17(depths: 90, 85,80,75,28,5m)
10/08/14	CTD_016	090	51°09.35'N 6°34.60'W	Bottles 1,4,5,6,8,9(depths: 91, 86 81,76,25,5m)
10/08/14	CTD_017	092	51°09.12'N 6°35.49'W	Bottles 10,13,14,15,17,18(depths: 88,83,78,73,32,5m)
10/08/14	CTD_018	094	51°08.98'N 6°35.83'W	Bottles 9,12,13,14,15,18,21(depths: 91,86,81,76,55,30,5m)
10/08/14	CTD_019	096	51°08.869'N 6°36.153'W	Bottles 1,4,5,6,7,9(depths: 90,85,80,75,21,5m)
10/08/14	CTD_020	098	51°08.8'N 6°36.31'W	Bottles 10,13,14,15,16,18(depths: 93,88,83,78,23,5m)
10/08/14	CTD_021	100	51°08.75'N 6°36.39'W	Bottles 19,22,23,24,1,3(depths: 93,88,83,78,27,5m)
10/08/14	CTD_022	102	51°08.74'N 6°36.39'W	Bottles 6,9,10,11,12,14(depths: 95,90,85,80,15,5m)
10/08/14	CTD_023	104	51°08.74'N 6°36.29'W	Bottles 1,4,5,6,7,10(depths: 95,90,85,80,18,5m)
10/08/14	CTD_024	106	51°08.74'N 6°36.238'W	Bottles 1,4,5,6,7,8(depths: 96,91,86,81,22,5m)
10/08/14	CTD_025	108	51°08.74'N 6°36.24'W	Bottles 2,4,5,6,7,9(depths: 99,90,85,80,23,5m)
10/08/14	CTD_026	110	51°08.7'N 6°36.24'W	Bottles 1,4,5,6,7,9(depths: 94,89,84,79,23,5m)
11/08/14	CTD_027	113	51°12.701'N 6°08.489'W	Bottles 1,4,7,10,13,16,19,22(depths: 95,70,50,31,28,25,15,5m)
12/08/14	CTD_029	135	51°08.91'N 6°36.24'W	Bottles 1,6,7,10,13,16,19,24(depths: 95,70,50,15,20,15,10,5m)
12/08/14	CTD_030	142	51°08.40'N 6°37.29'W	Bottles 1,3,5,8,9,13,15(depths: 92,75,55,35,24,20,5m)
13/08/14	CTD_033	159	51°12.574'N 6°03.353'W	Bottles 1,3,5,7,9,11,13,17,19(depths: 90,70,48,48,40,37,35,20,5m)

Preliminary Results

On 10th August we carried out hourly CTD's concentrating on the bottom 20 meters of the sediment, firing a bottle every 5 meters, plus taking the Chloro max and surface bottles.

The results below are for the concentrations of nutrients in the bottom waters, and show a tidal cycle with the water passing back and forth, but there was little or no evidence of sedimentary nutrient resuspension as had been postulated. Obviously, this is only a single experiment but implies little nutrient resuspension occurs in the late summer in the Celtic Sea.



Cruise Summary

The 5-channel autoanalyser worked very well throughout the cruise.

KANSO nutrient reference materials (Batch BU) run each day to check analyser integrity and analytical continuity from one day to the next. Very good continuity in sensitivity for all five channels was found, demonstrating excellent analytical performance.

Acknowledgements

Thank you to the Officers and Engineers of RRS *Discovery*, the NMF technicians and crew who made things work for us and kept them working, and of course the catering team for excellent food.

Determination of Oxygen Concentrations

Claire Mahaffey, University of Liverpool, UK

Methods

125ml optically clear glass oxygen bottles triple-rinsed with Milli-Q and stored full of Milli-Q. Each bottle pre-calibrated for volume and has unique identifying number on shoulder and on stopper. Oxygen samples drawn first from Niskin as soon as rosette secured on deck. Tygon tubing used to fill bottles from Niskin and bottle overflowed three times to ensure no bubbles. Temperature of each sample taken immediately then sample fixed with 1ml manganese sulphate (3M) and 1ml alkaline iodide, shaken vigorously for ~20 seconds. Samples re-shaken prior to storage approximately 15 minutes later.

Samples were stored upright under water in a dark 60L container until the precipitate had settled. Samples were analysed within 24 hours of collection. Prior to analysis 1ml sulphuric acid (10N) was added to each sample to dissolve the precipitate.

Samples were analysed for dissolved oxygen concentration onboard using the modified Winkler method (Carpenter, 1965) and a PC-controlled potentiometric titration system (Metrohm Titrand 888). Reagent blanks were run using 0.1N potassium iodate (1 aliquots) and sodium thiosulphate titrant (~ 0.18 N). Each of these was performed in triplicate (at minimum) prior to analysis of samples each day. Lab temperature monitored throughout analysis. Calculation of dissolved oxygen concentration was according to HOT protocol (website given below) and Grasshoff (1983). Samples were analysed to produce a dissolved oxygen concentration in $\mu\text{mol l}^{-1}$ and these values forwarded to the oxygen sensor calibration team for conversion to $\mu\text{mol kg}^{-1}$ and further processing.

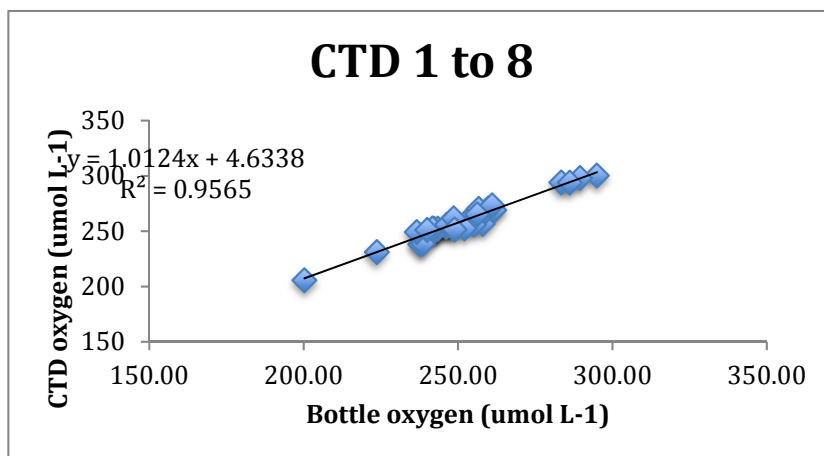
Samples were taken from 12 CTDs (Table 1). The coefficient of variation was typically better than 0.44% for triplicate analysis. Mean reagent blank was 0.0102 ± 0.005 mL over the course of the cruise and mean thiosulphate normality was 0.1732 ± 0.0008 N. Oxygen concentrations measured ranged from 200 $\mu\text{mol O}_2 \text{ l}^{-1}$ to 300 $\mu\text{mol O}_2 \text{ l}^{-1}$. Data to be submitted to BODC for conversion to $\mu\text{mol kg}^{-1}$ and calibration of the oxygen sensor on the CTD.

On CTD 13, it was noted that the oxygen sensor on the CTD giving highly variable readings. This is noted in the poor relationship the CTD oxygen and bottle oxygen from CTD9 to The oxygen sensor was cleaned just before CTD27, which improved the regression between the CTD oxygen and the bottle oxygen data although the slope (0.93, Figure 1c) was different to the slope estimated at the start of the cruise (1.02, Figure 1a).

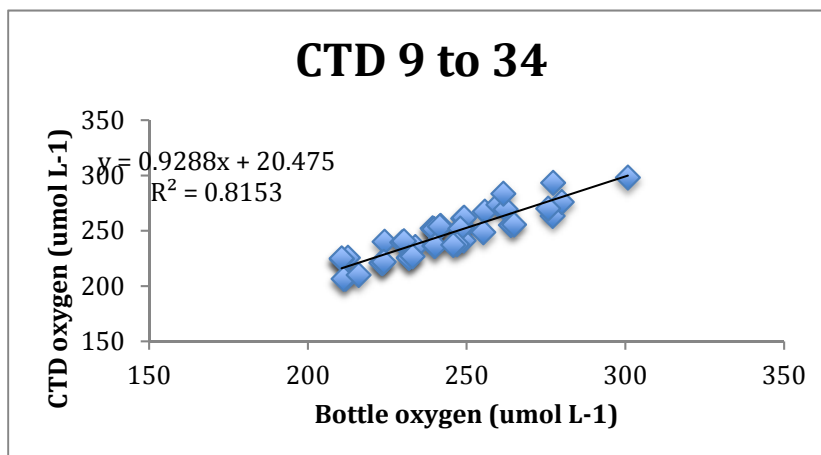
Table 1. List of rosette casts sampled for dissolved oxygen

Date	CTD no.	Lat	Long	Depths(m)
04/08/12	1	49° 48.8 N	5° 28.55 W	3 depths
05/08/14	6	49° 48.8 N	5° 28.66 W	7 depths
05/08/14	21	49° 22.3 N	8° 36.48 W	8 depths
06/08/14	30	48° 20.3 N	9° 43.63 W	9 depths
07/08/14	35	48° 34.2 N	9° 30.60 W	7 depths
08/08/14	56	50° 15.4 N	7° 44.62 W	8 depths
09/08/14	61	51° 08.3 N	6° 35.16 W	7 depths
09/08/14	78	51° 07.1 N	6° 37.52 W	8 depths
11/08/14	113			8 depths
12/08/14	142			6 depths
13/08/14	159			6 depths

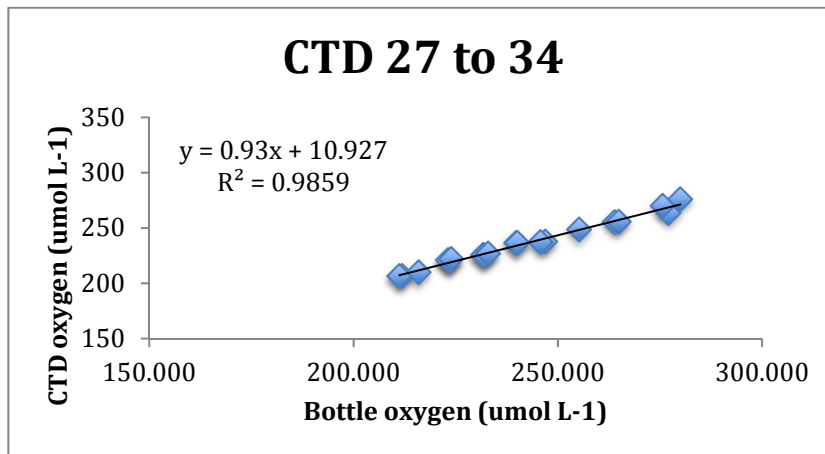
(a)



(b)



(c)



References

Carpenter, J.H. (1965). The Chesapeake Bay Institute Technique for the Winkler oxygen method. *Limnol. Oceanogr.*, **10**, 141–143.

Grasshoff, K. Ehrhardt, M, and K. Kremling (1983). *Methods of Seawater Analysis*. Grasshoff, Ehrhardt and Kremling, eds. Verlag Chemie GmbH. 419 pp.

<http://hahana.soest.hawaii.edu/hot/protocols/chap5.html>

Collection and Processing of Samples for Determination of Dissolved and Particulate Organic Matter

Clare Davis and Claire Mahaffey, University of Liverpool, UK

Dissolved Organic Nutrients

Samples collected from between 6 and 10 depths from the CTD and filtered through 47mm GF/F (combusted, Whatman, nominal pore size 0.7 μ m) and stored in acid-cleaned 175ml HDPE bottles at -20°C for later laboratory analysis to determine dissolved organic carbon (DOC), organic nitrogen (DON) and organic phosphorus (DOP) concentrations.

Dissolved Free and Total Hydrolysable Amino Acids

Samples collected from between 6 and 10 depths from the CTD and filtered through 47mm GF/F (combusted, Whatman, nominal pore size 0.7 μ m) and stored in 20ml muffled glass vials at -20°C, and then moved to the -80°C freezer for later laboratory analysis.

Coloured Dissolved Organic Matter (CDOM)

Samples were collected from between 6 and 10 depths from the CTD and underway system. Samples filtered through 47 mm GF/F (combusted, Whatman, nominal pore size 0.7 μ m) and then through 0.2 μ m Durapore filters. Samples kept in the dark and analysed on board using a Shimadzu UV-1650PC spectrophotometer and a Horiba Fluoromax-4 spectrofluorometer. Data will later be processed using PARAFAC by Nealy Carr (Sensors on Gliders Student) to determine the source and composition of CDOM.

Particulate Organic carbon, Particulate Organic Nitrogen and Particulate Phosphorus

Samples collected from between 6 and 10 depths from the CTD and marine snow catcher. For particulate carbon and nitrogen (PC/PN), 2L was filtered onto 25mm GF/F (combusted, Whatman, nominal pore size 0.7 μ m) on a plastic filter rig under <12 kPa vacuum pressure. For particulate phosphorus (PP), 1L was filtered onto 25mm GF/F (combusted and HCl acid washed, Whatman, nominal pore size 0.7 μ m) on a 3-port plastic filter rig under <12 kPa vacuum pressure. All filters were stored at -80°C for laboratory analysis.

Particulate Lipids and Particulate Amino Acids

Samples collected from between 6 and 10 depths from the CTD and marine snow catcher. For both lipids and amino acids, 3L was filtered onto 47 mm GF/F (combusted, Whatman, nominal pore size 0.7 μ m) on a 3-port glass filter rig under <12 kPa vacuum pressure. Filters were stored at -80°C for later laboratory analysis.

$\delta^{15}\text{N}$ of Particulate Nitrogen and Nitrate

Samples collected from between 6 and 10 depths from the CTD. Samples for the $\delta^{15}\text{N}$ and $\delta^{18}\text{O}$ of nitrate collected and stored in 60ml HDPE bottles (HCl acid washed) and stored unfiltered at -20°C for later

analysis. Samples for $\delta^{15}\text{N}$ -particulate nitrogen were collected by filtering 3L onto 25mm GF/F (combusted, Whatman, nominal pore size $0.7\mu\text{m}$) and stored at -80°C for later analysis.

Stand Alone Pump System (SAPS)

The SAPS was deployed four times to collect samples for PC/PN, PP, particulate lipids and particulate amino acids from two fractions: particles $>53\mu\text{m}$ and particles between $0.7 - 53\mu\text{m}$. Each time the SAPS was deployed to 50 m depth, to match that of snow catcher deployments made at similar times. The SAPS were programmed to pump for 1 hour once at that depth. Upon recovery, the $53\mu\text{m}$ mesh fraction was washed onto a 47mm GF/F (combusted, Whatman, nominal pore size $0.7\mu\text{m}$) which was stored at -80°C for later analysis. Below the mesh were two 27.3 cm diameter GF/Fs (combusted, Whatman, nominal pore size $0.7\mu\text{m}$) one was the sample GF/F and the second was stored as the blank GF/F, both were stored at -80°C for later analysis.

CTD Samples

Table 1. summarises the samples taken from the CTD for particulate carbon and nitrogen (PCPN), particulate phosphorus (PP), particulate lipids (LIPIDS), particulate amino acids (P-AA), dissolved and particulate δN^{15} (dN15), and dissolved organic nutrients including CDOM and amino acids (DOM).

Table 1: Summary of sample collection from the CTDs.

CTD #	Niskin #	PCPN	PP	LIPIDS	P-AA	dN15	DOM
2	3, 6, 9, 12, 18, 23	X	X	X	X		
3	2, 6, 7, 11, 16, 21 3, 5, 8, 12, 17, 20	X	X				X
4	2, 5, 7, 8, 11, 17, 20, 23	X	X			X	
5	2, 3, 4, 5, 10, 16, 18, 19, 23	X	X				X
6	1, 5, 8, 12, 16, 20, 23	X	X	X	X		
7	2, 4, 7, 10, 12, 16, 18, 22 1, 3, 6, 9, 11, 15, 17, 21	X	X				X
8	1, 5, 12, 15, 18, 21, 22	X	X			X	
9	1, 3, 7, 9, 12, 15, 18, 22						X
10	1, 4, 8, 12, 16, 20, 23	X	X	X	X		
11	2, 4, 6, 9, 16, 20, 24 1, 3, 5, 8, 15, 17, 19, 21	X	X				X
13	2, 5, 7, 11, 13, 17, 20, 23	X	X			X	
14	1, 4, 5, 6, 7, 8	X	X				X
15	9, 12, 13, 14, 16, 17	X	X				
16	1, 4, 5, 6, 8, 9	X	X				X
17	10, 13, 14, 15, 17, 18	X	X				

18	9, 12, 13, 14, 18, 21	X					X
19	1, 4, 5, 6, 7, 9	X	X				
20	10, 13, 14, 15, 16, 18	X	X				X
21	1, 3, 19, 22, 23, 24	X	X				
22	6, 9, 10, 11, 12, 14	X	X				X
23	1, 4, 5, 6, 7, 10	X	X				
24	1, 4, 5, 6, 7, 8	X	X				X
25	2, 4, 5, 6, 7, 9	X	X				
26	1, 4, 5, 6, 7, 9	X	X				X
27	1, 4, 7, 10, 13, 16, 19, 22						X
29	2, 6, 8, 11, 14, 18, 20, 24					X	
30	1, 3, 5, 9, 13, 15						X

Marine Snow Catcher: For the snow catchers, the filtering protocols were as stated above with the exception that for the fast sinking fraction (F3) the total tray contents were filtered rather than a volumetric measure.

Table 2. Summary of sample collection from the marine snow catcher

MSC #	Fraction	PCPN	PP	LIPIDS	P-AA
1	Susp (F1), Slow sinking (F2), Fast sinking (F3)	X	X	X	X
2	Susp (F1), Slow sinking (F2), Fast sinking (F3)	X	X	X	X
3	Susp (F1), Slow sinking (F2), Fast sinking (F3)	X	X	X	X
9	Susp (F1), Slow sinking (F2), Fast sinking (F3)	X	X	X	X
10	Susp (F1), Slow sinking (F2), Fast sinking (F3)	X	X		
11	Susp (F1), Slow sinking (F2), Fast sinking (F3)	X	X		
12	Susp (F1), Slow sinking (F2), Fast sinking (F3)	X	X	X	X
13	Susp (F1), Slow sinking (F2), Fast sinking (F3)	X	X		
14	Susp (F1), Slow sinking (F2), Fast sinking (F3)	X	X		
15	Susp (F1), Slow sinking (F2), Fast sinking (F3)	X	X		
16	Susp (F1), Slow sinking (F2), Fast sinking (F3)	X	X		
17	Susp (F1), Slow sinking (F2), Fast sinking (F3)	X	X		
18	Susp (F1), Slow sinking (F2), Fast sinking (F3)	X	X		
19	Susp (F1), Slow sinking (F2), Fast sinking (F3)	X	X		
20	Susp (F1), Slow sinking (F2), Fast sinking (F3)	X	X		
21	Susp (F1), Slow sinking (F2), Fast sinking (F3)	X	X		
22	Susp (F1), Slow sinking (F2), Fast sinking (F3)	X	X		

23	Susp (F1), Slow sinking (F2), Fast sinking (F3)	X	X		
24	Susp (F1), Slow sinking (F2), Fast sinking (F3)	X	X		

SAPs Deployments

SAPs were deployed on 5th August (SAPs 1 and 2), 7th August (SAPs 3) and 9th August (SAPs 4).

Table 3. Summary of SAPS deployments and volume filtered

SAPS	Depth	Deployment length (h)	Meter start	Meter End	Litres filtered
1	60 m	1	105457	105982	525
2	20 m	1	105982	106365	383
3	50m	1	106365	106962	597
4	50m	1	106962	107442	480

The Determination of Pelagic Nitrogen Regeneration Rates

Darren Clark, Plymouth Marine Laboratory, UK

Overview

Bacterial degradation of particulate and dissolved organic matter (P/DOM) simultaneously regenerates inorganic nutrients and renders the residual material of lower nutritional quality. Given sufficient time, the exposure of POM and DOM to a sufficiently broad range of microbes with their associated biochemical machinery renders P/DOM recalcitrant. This material represents a significant C-export flux. The preferential regeneration and retention of nutrients such as nitrogen and phosphorous during this process, generically termed the microbial carbon pump, sustains productivity of the shelf sea region.

During this program of research, pelagic nitrogen regeneration will be investigated. Specifically, the processes of NH_4^+ regeneration and nitrification will be examined. The former is primarily associated with the bacterially mediated degradation of organic molecules; if the C:N ratio of organic matter utilized by bacterial cells exceeds the cells C:N (i.e. cellular N-requirements) the excess is released as NH_4^+ . The latter is the two-stage oxidation of NH_4^+ to NO_2^- to NO_3^- , facilitated by specific clades of bacteria and Archaea. In combination, NH_4^+ regeneration and nitrification have the capacity to significantly influence concentration and composition of the dissolved inorganic nitrogen (DIN) pool, which sustains autotrophic primary production.

The aim of this research is to understand variability in N-regeneration processes, how rates relate to particle loading and how tidal re-suspension of benthic particles may influence exchange processes with the base of the water column.

Experiments

Large Marine Snow Catcher (LMSC) experiments: The regeneration of N associated with 3 particle fractions (suspended, slow and fast sinking) was determined during LMSC deployments below the photic zone (50m). The rates of NH_4^+ regeneration, NH_4^+ oxidation and NO_2^- oxidation measured on each fraction. Method details provided below. The volume of LMSC was 300L. Each quarter of the fast sinking particle tray represented the particle load from 75L of seawater. For deployments at station 008 and 040, 25% of the particle tray used for N-regeneration incubations. The remainder used for respiration measurements (Elena) and particle characterization (Emma). For station 060, 75% of the tray used with the remainder used for particle characterization.

Tidal study: The CTD was used to collect seawater samples from the base of the water column within close proximity to the seabed. Water was collected every 2 hours over a 12 hour period. An estimation of N-regeneration rates (NH_4^+ regeneration, NH_4^+ oxidation) in samples containing various degrees of tidally re-suspended material was undertaken.

Methods

The regeneration of DIN investigated using ^{15}N dilution methods (Clark et al 2006, 2007). The LMSC was used to collect seawater from a specific depth. When the aphotic zone was sampled the transparent viewing windows of the LMSC were covered to stop light intrusion. N-regeneration rates were determined in three particulate fractions (suspended particle (SP); slow sinking particle (SSP); fast sinking particle (FSP)). Following deployment and a 2 hour period of settling, 15L of SP seawater was collected from the LMSC. 1.5L of this water was added to each of 3 2.2L bottles containing either $^{15}\text{NO}_3^-$, $^{15}\text{NO}_2^-$ or $^{15}\text{NH}_4^+$. The ^{15}N addition estimated to provide a 20% enrichment of the DIN pool, based on recently determined nutrient concentration profiles. A further 4.0L of water containing SSP was collected from the MSC directly into bottles containing ^{15}N . FSP recovered in a tray from the LMSC, and in a constant temperature room under low intensity red light the particle tray screened for magnetic particles. FSP transferred to 2.2L bottles containing ^{15}N . One quarter of the total FSP load (equating to the FSP content of approximately 75L of seawater) added to each of 3 bottles (each representing one process). SP water used to dilute the FSP to a total volume of 1.5L. The 9 x 2.2. L bottles (3 processes, 3 particles fractions) were placed in a temperature controlled incubator for 30 minutes to ensure that the isotope was homogeneously distributed. Following this period, bottles were used to fill 1.0L incubation bottles and returned to the incubator for a period of 12 to 24 hours (experiments tested differing incubation times). The remaining ^{15}N amended seawater was filtered using 47mm GF/F. The filter was retained to enable a measure of particulate carbon and nitrogen content. The filtrate was used to derive the pre-incubation DIN concentration and isotopic enrichment by synthesizing indophenol from ammonium and sudan-1 from nitrite (nitrate is quantitatively reduced to nitrite prior to further analysis). Following the incubation period, samples filtered using GF/F. The filter was retained to enable an estimation of the particulate carbon and nitrogen content of the incubated sample. The filtrate was used to generate post-incubation samples for DIN concentration and isotopic enrichment.

Indophenol was synthesized in samples by adding the first reagent (4.7 g phenol and 0.32 g sodium nitroprusside in 200 mL Milli-Q water) in the proportion of 1 mL per 100 mL of sample volume, mixing the sample and leaving for 5 minutes. The second reagent (1.2 g sodium dichloro-isocyanurate and 2.8 g sodium hydroxide in 200 mL Milli-Q) added in the proportion of 1 mL per 100 mL sample volume, mixed and left for 5 hours at room temperature for indophenol development. Indophenol was collected by solid-phase extraction (SPE) as described below. Sudan-1 synthesized by adding the first reagent (0.8g of aniline sulphate in 200 mL 3M HCl) to samples in the proportion 0.5 mL per 100 mL sample volume. Samples were mixed and left for 5 minutes to homogenize after which sample pH was verified to be < 2.0 . Reagent 2 (24 g NaOH and 0.416 g 2-naphthol in 200 mL Milli Q) was added in the proportion 0.5 mL per 100 mL sample volume. Samples again were mixed, left for 5 minutes before sample pH was verified to be approximately 8.0. Sudan-1, the development of which was complete after 30 minutes of incubation at room temperature, collected by SPE as described below.

Deuterated internal standards added to samples immediately prior to SPE collection. Deuterated indophenol and deuterated sudan-1 were synthesised according to methods described previously (Clark et al. 2006; 2007). Standard solutions in methanol were prepared ($100 \text{ ng} \cdot \mu\text{L}^{-1}$) and the concentration verified against

analytical standard solutions (Sigma-Aldrich). Appropriate volumes of deuterated internal standards (i.e. comparable to samples size) were added to samples following acidification by citric acid and prior to SPE collection.

Indophenol and sudan-1 were collected by SPE using 6ml/500mg C18 cartridges (Biotage, UK) which were prepared for sample collection by first rinsing with 5ml methanol, followed by 5 mL Milli-Q water and 5 mL 0.22 µm filtered seawater. Prior to sample collection seawater samples were acidified with 1 M citric acid to a pH of 5.5, before collection by SPE under low vacuum (120 mmHg) at a flow rate of approximately 1 mL per minute without drying. Samples rinsed with 5ml 0.22 µm filtered seawater and 5ml Milli-Q water before being air dried under high vacuum (360 mmHg). Samples were stored frozen until further processing at the land based laboratory.

Table of Sampling Events

STNNBR	Date	Lat/Long	Gear	Depth	Process
008	5/8/14	49 23.18/8 37.13	LMSC	50m	NH ₄ ⁺ Reg
008	5/8/14	49 23.18/8 37.13	LMSC	50m	NH ₄ ⁺ Ox
008	5/8/14	49 23.18/8 37.13	LMSC	50m	NO ₂ ⁻ Ox
040	7/8/14	48 34.57/9 31.0	LMSC	50m	NH ₄ ⁺ Reg
040	7/8/14	48 34.57/9 31.0	LMSC	50m	NH ₄ ⁺ Ox
040	7/8/14	48 34.57/9 31.0	LMSC	50m	NO ₂ ⁻ Ox
060	9/8/14	51 08.26/6 35.16	LMSC	50m	NH ₄ ⁺ Reg
060	9/8/14	51 08.26/6 35.16	LMSC	50m	NH ₄ ⁺ Ox
060	9/8/14	51 08.26/6 35.16	LMSC	50m	NO ₂ ⁻ Ox
088	10/8/14	51 09.38/6 34.52	CTD(Nisk 2)	92m	NH ₄ ⁺ Reg/Ox
092	10/8/14	51 09.38/6 34.52	CTD(Nisk 2)	92m	NH ₄ ⁺ Reg/Ox
096	10/8/14	51 09.38/6 34.52	CTD(Nisk 10)	92m	NH ₄ ⁺ Reg/Ox
100	10/8/14	51 09.38/6 34.52	CTD(Nisk 11)	92m	NH ₄ ⁺ Reg/Ox
104	10/8/14	51 09.38/6 34.52	CTD(Nisk 7)	92m	NH ₄ ⁺ Reg/Ox
108	10/8/14	51 09.38/6 34.52	CTD(Nisk 2)	92m	NH ₄ ⁺ Reg/Ox
110	10/8/14	51 09.38/6 34.52	CTD(Nisk 2)	92m	NH ₄ ⁺ Reg/Ox

Status of samples and data availability

No data is available during the cruise. The samples are stored at -20°C in the form of solid-phase extraction cartridges and GF/F filters to be analysed at the land-based laboratory. The former will be used for isotope dilution studies and the later for quantifying the carbon and nitrogen content of incubated samples. Analysis will take approximately 6 weeks, after which a high quality data set is expected to be delivered.

Modifications to be made for the SSB cruise program.

- LMSC deployments for N-regeneration studies undertaken at night, removing the risk that samples are exposed to sunlight

- 75% of the particle tray from LMSC deployments will be used for N-regeneration incubations. An incubation period of 24 hours used
- The full programme will include N-assimilation rate determinations. This will include urea assimilation

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Shelf-sea Gross and Net Production Estimates from Triple Oxygen Isotopes and Oxygen-argon Ratios in relation with Phytoplankton Physiology

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Objectives

1. Infer spatial variations of net (N) and gross (G) O_2 production rates from O_2/Ar [N (O_2/Ar)] and triple oxygen isotopes [G (^{17}O)] in the Celtic Sea.
2. Derive 24 h-in situ production rates from diurnal changes at process stations
3. Calculate seasonally integrated production estimates from cruise-to-cruise changes
4. Compare G (^{17}O) with FRRF-based physiological turnover and CO_2 fixation rates
5. Use statistical tools to relate N and G to production estimates based on ^{15}N - and ^{14}C -uptake, respiration rates, light availability, nutrient supply, community structure and other SSB consortium data products

Introduction

In order to increase the resolution of dynamic waters such as shelf seas, continuous underway measurement systems are a good choice.

Membrane inlet mass spectrometry is a technique invented by Hoch and Kok in 1963. This technique permits the sampling of dissolved gases from a liquid phase. The principle is a semipermeable membrane that allows dissolved gases pass through but not the liquid into the mass spectrometer flying tube. This technique was considered very sensitive (Hoch and Kok, 1963) but nowadays, even if modern MIMS have high sensitivity (Beckmann et al., 2009) these instruments lack the ultra-high precision of IRMS. The advantages of the MIMS are several with the exception of the precision. These can be mounted onboard which permit the analysis of several dissolved gases of seawater in situ and continuously. Phytoplankton photosynthesis and respiration understandings can be achieved from the analysis of stable isotopes distribution of certain gases or to obtaining chemical exchange rates (Beckmann et al., 2009). This is also a very simple way to analyze volatile gases, do not require exhaustive preparation of material for sampling nor the use of chemicals, and data is recorded directly in the computer without the need of post analysis in the laboratory.

The dissolved O_2 in seawater gives an estimation of the NCP. Physical process such as variation in temperature and pressure, transport fluxes, diffusion and bubble injection also changes the amount of dissolved O_2 in seawater. Now is clear that we need a tracer that separates oxygen produced biologically from the one added or removed from physical processes. Argon does not react during photosynthesis or respiration and have similar solubility and diffusivity than O_2 . Variation in O_2 concentration due to biological production can be separated from physical forces using the $\Delta O_2/Ar$ ratio.

Craig and Hayward (1987) were the first ones describing a technique for using ΔO_2 and Ar differences to determine NCP. The equation that is now used is $\Delta O_2/Ar$ ratio, and defined as follows in Eq. (1).

$$\Delta O_2/Ar = [c(O_2)/c(Ar)]/[c_{sat}(O_2)/c_{sat}(Ar)]-1 \quad (1)$$

Where c is the dissolved gas concentration (mol m^{-3}) and c_{sat} is the saturation concentration at known temperature, pressure and salinity (Kaiser, 2005).

In addition to the underway measurements, discrete samples were taken for calibration purposes and to measure the $^{17}\text{O}/^{16}\text{O}$ and $^{18}\text{O}/^{16}\text{O}$ isotope ratio analysis of dissolved oxygen. Triple oxygen isotope measurements combined with O_2/Ar data can be used to estimate the ratio of net community production to gross production and the ratio of gas exchange to gross production. Again, in combination with suitable wind-speed gas-exchange parameterizations this can be used to estimate gross production over large regional scales at timescales of weeks to months.

Methodology

Continuous measurements of dissolved N_2 , O_2 , and Ar made by MIMS on board RRS *Discovery*. The ship's Underway Sampling System was used to pump water through a tubular Teflon AF membrane (*Random Technologies*). The membrane connected to the vacuum of a quadrupole mass spectrometer (*Pfeiffer Vacuum Prisma*). The intake of the underway sampling system is located at the bow at a nominal depth of 5 m. The water from the underway sampling system passed through an open bottle at several litres per minute to remove macroscopic bubbles and to avoid pressure bursts. A flow of about 45 ml/min was continuously pumped from the bucket through the membrane, using a gear pump (*Micropump*). In order to reduce O_2/Ar variations due to temperature effects and water vapour pressure variations, the exchange chamber with the membrane was held at a constant temperature of 12°C (5 to 10°C below the sea surface temperature, to avoid temperature-induced super-saturations and subsequent bubble formation). The flight tube was in a thermally insulated box maintained at 50°C.

In addition to the continuous underway MIMS measurements, I also analysed CTD samples in order to characterize the depth profile of the O_2/Ar ratio in regions of the Celtic Sea.

The O_2/Ar ratio measurements will be calibrated with discrete water samples taken from the same seawater outlet as used for the MIMS measurements. 200cm³ samples were drawn into pre-evacuated glass flasks poisoned with 7mg $HgCl_2$ [Quay *et al.*, 1993]. These samples will be later analyzed with an isotope ratio mass spectrometer (IRMS, *Thermo Finnigan*) for their dissolved O_2/Ar ratios and the oxygen triple isotope composition relative to air [Hendricks *et al.*, 2004]. Raw O_2/Ar ion current ratio measurements made every 10 to 20 s and had a short-term stability of 0.05%.

O_2 concentrations were also measured continuously with an optode (*Aanderaa* model 3830, serial no. 241), resolution of 10 second. The measurements were from the open bottle connected to the underway sampling system that I have used to measure the O_2/Ar ratios.

Discrete samples

The CTD profile has shown a stratified water column during all the cruise sampling. The mixed layer was between 15 to 40metres deep and the euphotic zone was always around 5metres deeper. Peaks of chlorophyll maximum and oxygen were mostly found in the below the bottom of the mixed layer and in the euphotic zone. The following samples collected:

<u>CTD</u>	<u>Latitude N</u>	<u>Longitude W</u>	<u>Start date</u>	<u>Start time (GMT)</u>	<u>Niskin Bottle</u>	<u>Depth</u>	<u>500ml bottle</u>	<u>Ev. Bottle</u>
1	49 48 80	005 28 66	04/08/2014	11:20:00	19	5	24	
					15	17	25	945
					11	25	27	991
					9	35	28	
					8	50	29	
					6	65	31	
2	49 23 194	08 37 127	05/08/2014	5:10:00	2	80	36	
					22	5	24	
					17	20	25	
					14	30	27	956
								997
								998
4	49 22 311	08 36 465	05/08/2014	16:08:00	11	45	28	964
								952
								961
					8	60	29	
					5	100	31	
					24	5	24	
5	48 20 30	009 43 62	06/08/2014	8:24:00	21	20	25	
					15	40	27	983
								946
								960
					9	45	28	963
								967
5	48 20 30	009 43 62	06/08/2014	8:24:00	7	60	29	
					22	5	24	
					16	25	25	976

6	48 34 193	009 30 616	07/08/2014	6:05:00	12	35	27	947
					9	50	28	
					6	75	29	
					2	1000	31	
					24	5	24	
					21	15	25	
					18	35	27	957
								971
								958
					10	40	28	943
7	48 34 59	009 30 92	07/08/2014	11:09:00				972
								851
					9	50	29	
					6	120	31	
					24	5	24	
					19	20	25	
					14	42	27	970
								944
								968
					8	55	28	936
8	48 34 270	009 30 276	07/08/2014	15:00:00				980
								962
					6	90	29	
					5	120	31	
					23	5	24	
					20	15	25	166
					17	29	27	975
								949
								990
					14	40	28	950
9	50 15.483	007 44.611	08/08/2014	11:04:00	11	60	29	965
					9	90	31	
					20	5	24	
					19	20	25	175
								940
								996
					13	34	27	995
			984					

								994
					8	45	28	959
					4	60	29	
					2	93	31	
10	51 08.265	06 35.149	09/08/2014	6:09:00	24	5	24	
					18	20	25	96
								124
								62
					17	30	27	
					13	35	28	101
								183
								77
					9	50	29	
					6	60	31	
					3	85	36	
					11	35	phyto	10L
11	51 7.248	6 37.283	09/08/2014	10:40:00	24	5	24	
					18	28	25	113
								79
								73
					15	34	27	109
								202
								12
					10	36	28	
					7	50	29	
					3	70	31	
12	51 7.097	6 37.498	09/08/2014	16:05:00	24	5	24	
					21	15	25	108
								5
								78
					18	25	27	103
								90
								82
					15	37	28	
					9	48	29	
					6	65	31	
15	51 9.41	6 34.252	10/08/2014	8:23:00	15	30	24	84
16	51 9.35	6 34.600	10/08/2014	9:16:00	7	31	24	74

17	51 9.17	6 3536	10/08/2014	10:13:00	16	36	24	106
18	51 8.989	6 35.833	10/08/2014	11:22:00	17	30	24	93
19	51 8.869	6 36.153	10/08/2014	12:18:00	8	17	24	105
20	51 8.80	6 36.31	10/08/2014	13:14:00	17	16.3	24	100
21	51 8.75	6 3639	10/08/2014	14:17:00	2	17	24	110
22	51 8 747	6 36.39	10/08/2014	15:25:00	13	9	24	88
23	51 08.742	6 36.29	10/08/2014	16:24:00	22	18	24	97
24								
25	51 8 74	6 36.24	10/08/2014	18:20:00	8	15	24	94
26	51 8 74	6 36.24	10/08/2014	19:24:00	8	15	24	992
27	51 12.701	6 8.489	11/08/2014	11:00:00	23	5	24	
					20	15	36	942
					17	25	25	
					14	28	27	941
					11	31	28	
					8	50	29	
					5	70	31	
29	51 8.91	6 36.24	12/08/2014	7:00:00	4	70	24	
							4	
					5	70	25	
					16	15	27	
							20	
					17	15	28	
					22	5	29	
							36	
					23	5	31	
31	51 8.218	6 37.771	12/08/2014	13:44:00	4	21	4	
					4	21	20	
					4	21	36	
					4	21	31	
					4	21	28	
					4	21	25	

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Phytoplankton Community Composition and Marine Snow Catcher Measurements focusing on the Chlorophyll Maximum

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Introduction

In seasonally stratified temperate coastal and shelf seas, a mid-water chlorophyll maximum, 'thin layer', is often detectable just below the thermocline, with associated increased abundances of phytoplankton cells. The relative importance of these summer subsurface chlorophyll maxima in relation to export production however, has not been previously investigated. Therefore, the main aim for the cruise was to sample these chlorophyll maxima in order for later establishment of their potential for export of organic carbon.

Research Approach

Two sampling devices used:-

1. A CTD mounted on a Niskin rosette system
2. A small Marine Snow Catcher

Samples from Niskin bottles and from the Marine Snow Catcher processed as follows: -

CTD Niskin Samples

Niskin bottles on the CTD rosette system sampled when a chlorophyll maximum was present. Since phytoplankton community composition is a key factor that influences export production, lugol's and glutaraldehyde samples were collected for 4-7 depths (spanning depth to surface, and on many occasions spanning the chlorophyll maximum) for many of the cruise CTD casts, along with occasional size fractionated chlorophyll (total, >10 μ m and >50 μ m) and HPLC samples. Therefore, phytoplankton community composition of the chlorophyll maxima, bottom mixed layer and upper mixed layer could later be assessed.

Marine Snow Catcher Samples

Three small marine snow catcher deployments to just below the chlorophyll maximum and three paired small marine snow catcher deployments to just below the chlorophyll maximum and in the upper mixed layer conducted during the cruise, with the suspended, slow sinking and fast settling fractions being analysed for all deployments. For the suspended and slow sinking fractions POC, HPLC and lugol's samples collected. While the fast sinking fraction was photographed using an imaging rig to allow for later determination of the POC content of the fraction, and then particle setting experiments were conducted to allow for later determination of the sinking rate of the fast sinking fraction. Additionally, 18-20 randomly picked aggregates were placed on a GF/F filter to allow for later pigment analysis of the fast sinking fraction using the HPLC technique.

Techniques Employed

- Lugol's preservation - 1ml of lugol's iodine added to 50ml of sample
- Glutaraldehyde preservation - 50 μ l of 50% glutaraldehyde added to 10ml of sample
- Size fractionation - 50ml of sample filtered through appropriately sized mesh or track-etched membranes, before being filtered through a 25mm dia. GF/F filter (filtration performed using a syringe pump)
- HPLC sample collection - 1-2L of sample filtered through a 25mm dia. GF/F filter using a filtration rig
- POC sample collection - 1-2L of sample filtered through a pre-combusted 25mm dia. GF/F filter using a filtration rig
- Settling experiments - 2 sinking times recorded as individual particles passed two discrete points within a 2L measuring cylinder filled with suspended fraction seawater

Measurements

Date	Station	Event	Depths (m)	Measurements
05/08	011	CTD cast 3	138.7	Lugol's and glutaraldehyde
			101.4	
			51.8	
			35.4	
			21.1	
			5.8	
05/08	021	CTD cast 4	136.4	Lugol's and glutaraldehyde
			100.8	
			45.8	
			40.4	
			30.8	
			5.7	
06/08	030	CTD cast 5	125.0	Lugol's, Glutaraldehyde and HPLC on all depth samples, with size fractionated chlorophyll measurements just on 26.6m and 25.4m
			50.0	
			26.6	
			25.4	
			5.8	
06/08	031	MSC 4	~35.0	HPLC, POC and Lugol's samples taken for suspended and slow sinking fraction. Particle settling expt.s on fast sinking fraction, with HPLC aggregate sample also taken
07/08	035	CTD cast 6	120.4	Lugol's and glutaraldehyde

			51.2	
			40.4	
			35.4	
			15.8	
			5.6	
07/08	046	CTD cast 8	~120.0	
			~90.0	
			~60.0	
			~40.0	
			~29.0	
			~5.0	
08/08	056	CTD cast 9	61.2	Lugol's, Glutaraldehyde and HPLC on all depth samples, with size fractionated chlorophyll measurements just on 37.0m, 35.0m and 33.7m
			46.1	
			37.0	
			35.0	
			33.7	
			6.1	
08/08	057	MSC 8	~40.0	HPLC, POC and Lugol's samples taken for suspended and slow sinking fraction. Particle settling expt.s on fast sinking fraction, with HPLC aggregate sample also taken
09/08	061	CTD cast 10	~85.0	Lugol's and glutaraldehyde on all depth samples, with HPLC on 35.6m
			61.6	
			51.9	
			35.6	
			~30.0	
			6.4	
09/08	071	CTD cast 11	~70.0	Lugol's, glutaraldehyde and HPLC on all depth samples, with size fractionated chlorophyll measurements just on 36m, 34m and 28.2m
			~50.0	
			36.0	
			34.0	
			28.2	
			6.0	
09/08	072	MSC 13	~40.0	HPLC, POC and Lugol's samples taken for suspended and slow sinking fraction. Particle settling expt.s on fast sinking fraction, with HPLC aggregate sample also taken

09/08	078	CTD cast 13	~65.0	Lugol's, glutaraldehyde and HPLC
			39.0	
			37.4	
			25.4	
			~5.0	
11/08	112	CTD cast 27	50.7	Lugol's, glutaraldehyde and HPLC (+ chlorophyll to be analysed by Emma)
			31.8	
			29.0	
			25.8	
			~5.0	
11/08	116	CTD cast 28	60.8	Lugol's, glutaraldehyde and HPLC (+ chlorophyll to be analysed by Emma)
			34.6	
			33.5	
			32.1	
			5.5	
11/08	117	MSC 27	~40.0	HPLC, POC and Lugol's samples taken for suspended and slow sinking fraction. Particle settling expt.s on fast sinking fraction, with HPLC aggregate sample also taken
11/08	118	MSC 28	~20.0	HPLC, POC and Lugol's samples taken for suspended and slow sinking fraction. Particle settling expt.s on fast sinking fraction, with HPLC aggregate sample also taken
12/08	142	CTD cast 30	56.0	Lugol's and glutaraldehyde (+ chlorophyll to be analysed by Emma)
			25.6	
			21.5	
			6.7	
12/08	146	CTD cast 31	61.8	Lugol's, glutaraldehyde, HPLC, chlorophyll and nutrients
			21.4	
			17.7	
			18.2	
			6.7	
12/08	147	MSC 35	~30.0	HPLC, POC and Lugol's samples taken for suspended and slow sinking fraction. Particle settling expt.s on fast sinking fraction, with HPLC aggregate sample also taken
12/08	148	MSC 36	~15.0	HPLC, POC and Lugol's samples taken for

				suspended and slow sinking fraction. Particle settling expt.s on fast sinking fraction, with HPLC aggregate sample also taken
12/08	150	CTD cast 32	50.7	Lugol's, glutaraldehyde, HPLC, chlorophyll and nutrients (no HPLC on 14.0m)
			26.0	
			23.0	
			20.2	
			17.5	
			14.0	
			6.6	
12/08	151	MSC 37	~30.0	HPLC, POC and Lugol's samples taken for suspended and slow sinking fraction. Particle settling expt.s on fast sinking fraction, with HPLC aggregate sample also taken
12/08	152	MSC 38	~15.0	HPLC, POC and Lugol's samples taken for suspended and slow sinking fraction. Particle settling expt.s on fast sinking fraction, with HPLC aggregate sample also taken

Measurements of Community and Bacterial Respiration by Changes in O₂ Concentration after 24 Hours Incubation, *in vivo* INT Reduction Capacity Method and Continuous Oxygen Decrease using Oxygen Optodes

E Elena Garcia-Martin, University of East Anglia and Michelle Barnett, University of Southampton

The aims of this work were:

1. To determine the variability of the organic carbon remineralisation (community and bacterial respiration, CR and BR) during a tidal period and study the effect of the material re-suspended by tides on the respiration.
2. To settle the protocol to measure community and bacterial respiration with *in vivo* INT reduction capacity method (ivINT method), in order to be able to estimate accurate bacterial growth efficiencies of particle attached bacteria.
3. To quantify community and bacterial respiration of the three fractions of the Marine Snow Catcher (suspended, slow sinking and fast sinking) above and below the thermocline with Winkler technique and ivINT method (once established the protocol).
4. To log and quantify continuously the respiration of fast and suspended particles with an oxygen optode.

Sampling and analytical methodology

Seawater collected directly from Niskin bottles from three morning CTD casts (Table 1) from three depths in 10L carboys. The sampling depths were above & below the thermocline (matching the Marine Snow Catcher deployment) and the deep chlorophyll maximum. Each carboy subsampled for measuring community respiration by *in vitro* changes of dissolved oxygen concentration, community and bacterial respiration by the size-fractionated *in vivo* INT reduction capacity method (see below).

Water samples from the suspended, slow and fast sinking fractions collected from the Marine Snow Catcher at four stations (Table 2). Suspended material and slow sinking collected in 2-5L carboys and transported to a dark room for subsequent subsampling and analysis of community and bacterial respiration, as outlined below. The fast sinking material was taken from one of the quarters of the tray in dark conditions. Special care taken at all moments to prevent the exposure of the samples to light, and a red light was used while handling the samples (Figure 1). The fast sinking particles gently siphoned with a pipette into a bottle and subsampled from here to the different methodologies. As the water volume was not enough for the different techniques, dilutions of 10:1 and 1:1 (suspended: fast) were applied.

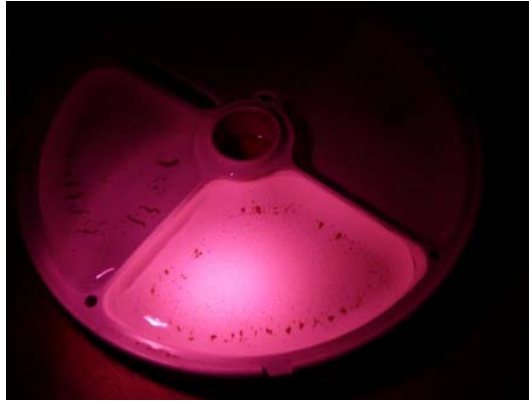


Figure 1. Tray of a large Marine Snow Catcher and the fast sinking particles

Community respiration by in vitro changes of dissolved oxygen concentration

CR measured by monitoring changes in oxygen concentrations after 24h dark bottle incubations. Dissolved oxygen concentration measured by automated precision Winkler titration performed with a Metrohm 765 Titrino titrator, utilising a photometric endpoint (Carritt & Carpenter, 1966).

Six gravimetrically calibrated 60 ml glass Winkler bottles were carefully filled with water from each depth. Water was allowed to overflow during the filling, and special care was taken to prevent air bubble formation in the silicone tube. Three bottles were fixed at start of the incubation (“zero”) with 0.5 ml of sulphate manganese and 0.5 ml of a solution of sodium iodine/sodium hydroxide. The other three bottles were placed in a water temperature controlled incubators inside the CT room for 24 hours. Bottles removed from the incubators after the 24 hours and fixed as the “zero”. All bottles were analysed within the next 24 hours. The concentrations of the thiosulphate used were 0.1 and 0.12 N. Thiosulphate calibrated every day before the analysis of the samples.

Community respiration calculated from the difference in oxygen concentration between the means at time zero and at 24 hours dark incubation.

***In vivo* community and bacterial respiration (CR_{INT} and BR_{INT}) by enzymatic assay**

Four 50-200ml amber glass bottles filled with seawater from each 10L carboy from the CTD and seawater from the different fractions from the Marine Snow Catcher. One replicate immediately fixed by adding formaldehyde (2% w/v final concentration) and used as a killed control. Twenty minutes later all four replicates were inoculated with a sterile solution of 7.9 mM 2-(*p*-iodophenyl)-3-(*p*-nitrophenyl)-5-phenyl tetrazolium salt (INT) to give a final concentration of 0.8 mM. The solution was freshly prepared for each experiment using Milli-Q water. Samples incubated in the same temperature controlled water bath as the dissolved oxygen bottles for 1-2 hours and then fixed by adding formaldehyde, as for the killed control. After 20 minutes, samples were put inside an ultrasound bath for one minute and then they were sequentially filtered through 0.8 and 0.2 μ m pore size polycarbonate filters, air-dried, and stored frozen in 1.5ml Cryovials at -20°C until further processing (one or two days later). The CR_{INT} (i.e. the sum of respiration of the >0.8 μ m and 0.2-0.8 μ m fractions) and BR_{INT} (considered as the respiration of the 0.2-0.8 μ m fraction) were measured following Martínez-García et al. (2009) with a Helios spectrophotometer.

This is the first time that this novel technique was applied to sinking particles and several tests were carried out in order to know the optimal sonication time to detach the bacteria from the particles and a time-course experiment in order to know the optimal incubation time that these samples should be incubated.

Optimal incubation time test

Seventeen samples of 50ml from the fast sinking particles from the Marine Snow Catcher and from the CTD were collected and dispensed into glass bottles. Incubations undertaken in the dark for 0, 0.5, 1, 1.5, 2 and 4 at in situ temperature. Optimal incubation time was considered as the time-period, prior to saturation of the formazan concentration, during which the relationship between concentration versus time remained linear. The optimal incubation time found to be <2 h and this was adopted as the maximum incubation time for the INT reduction assay.

Optimal sonication time test

Five samples of 40ml water (10:1, suspended: fast sinking particles) collected in glass bottles and fixed with glutaraldehyde. Bottles put inside an ultrasound bath for 0, 5, 10, 30 and 60 seconds. After the sonication time, samples taken for DAPI counts (see McNeill report).

Dilution test

A dilution test applied in order to test if the dilution applied to the fast sinking particles affected the respiration rates measured with the Winkler and ivINT technique. The dilution tested were 1:1, 10:1 (suspended: fast sinking) and non-diluted. Respiration estimated with changes in oxygen concentration and in vivo INT reduction methods. The sampling procedure was the same as described above but the four quarters of the tray from the small Marine Snow Catchers used.

There are no data from the fast particles from the Winkler technique as the fast sinking particles were full of sand that interfered with the photometric endpoint detector.

Continuous monitoring of in vitro oxygen evolution

Changes in oxygen concentration were measured continuously with three optode systems (YSI ProODO). Prior to each experiment, all the sensors were air calibrated simultaneously. 100ml seawater sample from the suspended water collected and filtered by 0.2 μm pore size polycarbonate filters. Samples from the suspended, fast sinking fraction and the filtered suspended water of the deep Marine Snow Catcher, were taken into 50ml glass bottles and left inside the water bath system to acclimate during 0.5-1 hour. This was done as the samples experienced a temperature increase during the settle time on deck (2 hours, see Cavan et al. report for the deployment and procedure with the Marine Snow Catcher). Incubation performed at the in situ temperature conditions ± 0.5 °C inside a dark water bath (Figure 2). The filtered water used as a background for abiotic changes in oxygen concentration associated to any temperature changes that the samples could have experienced during the incubation inside the water bath. After one hour of acclimation, 5-6 ml subsamples taken and put inside the YSI ProODO glass chambers. The chambers sealed to the probe

with Parafilm®. Oxygen concentration recorded every minute during c.a. 24 hours in a chart recorded. Oxygen consumption rates of the fast sinking and suspended material were determined as the slope of the oxygen concentration decrease as a function of time.



Figure 2. YSI ProODO optodes Deployment and the WaterBath Used

Preliminary Results

3 vertical profiles of three depths were sampled for community and bacterial respiration rates (Winkler and ivINT method).

4 incubations for continuous oxygen consumption (ProODO YSI optodes) were run with fast sinking particles from the Marine Snow catcher.

4 Marine Snow Catchers were sampled to calculate the carbon remineralization rates of the different fractions above and below the thermocline.

1 tidal effect experiment was performed sampling every two hours at 5metres under the surface and 5metres above the seabed for in vitro oxygen consumption and at 5metres above the seabed for in vivo INT reduction method.

2 time-course experiments for the in vivo INT reduction capacity method were done, one with samples from the CTD and the other with the slow sinking fraction of a Marine Snow Catcher samples.

1 sonication time-course experiment was performed in order to know the optimal sonication time to detach as many bacteria as possible without damaging the cells.

1 dilution test was performed in order to check if the dilution of the fast sinking particles with suspended water from the same depth affect the re-mineralization rates.

Respiration analyses all performed on board, data processed on return.

References

Carritt, D.E. and Carpenter, J.H., 1966. Comparison and evaluation of currently employed modifications of the Winkler method for determining dissolved oxygen in seawater; a NASCO Report. *Journal of Marine Research*, 24: 286-319.

Martínez-García, S., Fernández, E., Aranguren-Gassis, M., Teira, E., 2009. *In vivo* electron transport system activity: a method to estimate respiration in natural marine microbial planktonic communities. *Limnology and Oceanography Methods* 7, 459-469.

Table 1. List of collected water samples for measurements of respiration from CTD

Gear Code	St N	Date	Time	Site	Latitude	Longitude	Niskin	Depth (m)	Variable/Test
CTD	1	04/08/2014	10:21	Test south of Lizard	49 48.8 N	5 28.66 W	19	5	Optimum incubation time
CTD	6	05/08/2014	05:10	Candyfloss	49 23.18 N	8 37.13 W	17, 11, 8	20, 45, 60	
CTD	35	07/08/2014	06:04	Shelf break	48 34.2 N	9 30.6 W	21, 17, 9	15, 35, 50	
CTD	56	08/08/2014	11:05		50 15.48 N	7 44.62 W			Optode test
CTD	61	09/08/2014	06:08	Celtic Deep	51 08.26N	6 35.16 W	21, 13, 10	10, 35, 50	
CTD	86	10/08/2014	06:58	Celtic Deep	51 09.42N	6 34.28 W	8, 3	5, 92	
CTD	90	10/08/2014	08:59	Celtic Deep	51 09.34N	6 34.65 W	9, 3	5, 92	
CTD	94	10/08/2014	11:03	Celtic Deep	51 08.99 N	6 35.83 W	21, 11	5, 91	
CTD	98	10/08/2014	12:59	Celtic Deep	51 08.80 N	6 36.31 W	18, 12	5, 90	
CTD	102	10/08/2014	15:08	Celtic Deep	51 08.75 N	6 36.39 W	14, 8	5, 95	
CTD	106	10/08/2014	17:00	Celtic Deep	51 08.74 N	6 36.24 W	9, 3	5, 96	
CTD	110	10/08/2014	19:06	Celtic Deep	51 08.74 N	6 36.24 W	10, 3	5, 94	

Table 2. List of collected water samples for measurements of respiration from the Marine Snow Catchers

Gear Code	St. N	Date	Time	Site	Latitude	Longitude	Depth (m)	Variable/Test	Notes
LMSC	4	04/08/2014	14:27	Test south of Lizard	49 48.71 N	5 28.69 W	40	Optimum incubation time	1 quarter of the fast sinking tray
LMSC	7	05/08/2014	06:05	Candyfloss	49 23.18 N	8 37.13 W	20		1 quarter of the fast sinking tray
LMSC	8	05/08/2014	07:25	Candyfloss	49 23.18 N	8 37.13 W	60		1 quarter of the fast sinking tray
LMSC	12	05/08/2014	12:04	Candyfloss	49 23 N	8 36.6 W	100		1 quarter of the fast sinking tray
LMSC	34	07/08/2014	05:48	Shelf break	48 34.2 N	9 30.6 W	50		1 quarter of the fast sinking tray
LMSC	40	07/08/2014	08:48	Shelf break	48 34.57 N	9 31.0 W	10		
LMSC	62	09/08/2014	06:42	Celtic Deep	51 08.26N	6 35.16 W	50		3 quarter of the fast sinking tray
LMSC	70	09/08/2014	10:22	Celtic Deep	51 07.29N	6 37.22 W	10		2 quarter of the fast sinking tray
SMSC	136	12/08/2014	07:42	Celtic Deep	51 08.91N	6 36.24W	100		whole tray fast sinking particles
SMSC	137	12/08/2014	08:02	Celtic Deep	51 08.91N	6 36.24W	100		whole tray fast sinking particles

Bacterial Production Measurements

Sharon McNeill, Scottish Association for Marine Science

Introduction

Radio-labelled leucine methods were used to determine bacterial production in the Celtic Sea. Water column and Marine Snow Catcher samples chosen to correspond to respiration studies. A full list of bacterial production samples taken and analysed on board shown in Table 1.

Method

Leucine

Water samples were collected from the CTD in acid washed polycarbonate bottles then incubated for bacterial production, samples were also taken for flow cytometer and DAPI counts these were frozen at -80°C to be analysed back at SAMS. For bacterial production aliquots of 10ul leucine working solution (0.01 MBq ml⁻¹) were pipetted into each 2ml sterile centrifuge tube then additions of 1.6ml sample added this was carried out in the radioisotope container. For each depth, two samples in duplicate were run for T0, T1, T2 and T3 then incubated in a cool box in the CT room at above & below thermocline temperatures. Samples fixed with 80ul of 20% paraformaldehyde (giving a final concentration of 1%). Samples transferred to the radiochemistry container for processing, 25mm GFF and 0.2um polycarbonate filters presoaked in 1mM non-labelled leucine in separate petri dishes, placed on the 25mm filter rig with the GFF as a backing filter. Additions of 2ml of deionised water added onto the filter unit then the sample pipetted into each filter holder. Both samples at each time point combined and filtered as one. To remove the remaining sample the tube was rinsed with deionised water. The 0.2um polycarbonate filter was placed into a scintillation vial and dried overnight in the fume-hood, 4ml Optiphase Hi-Safe II scintillant was added and samples read in the scintillation counter after 24 hours. Marine Snow Catcher samples were analysed on three fractions, suspended, slow and fast sinking using the method describe above and also on 5ml sample volumes at 1:10, 1:5 and 1:1 dilutions with fast and suspended fractions. Marine snow catcher fast fractions samples taken from a ¼ tray approx. 40ml of the 200ml shared with Elena.

Calibration experiment - Leucine

Three replicate water column samples A, B and C were prepared into a 1litre polycarbonate bottle, 900ml of each filtered through a 0.2um filter vacuum cap with 100ml unfiltered making up the volume. Each replicate sampled at T0, T6, T12, T18 and T24 for leucine, bacterial count for flow cytometer and dapi slide prep. Samples incubated in a screened deck tank then then processed as water column methods for leucine.

Sonication experiment- Dapi

A snow catcher sample was taken from fast particulate material on the first day for a sonication trial. Samples were fixed in 1% glutaraldehyde and sonicated in duplicate for 0.5, 10, 30 & 60 seconds. Samples stained with DAPI at 25µl per 5ml sample and left to stain for a maximum of 5 minutes. The sample was

filtered onto a 0.2µm black polycarbonate filter with a 0.8µm cellulose nitrate as a backing filter. The filter placed on a microscope slide and frozen at -20°C until ready to enumerate.

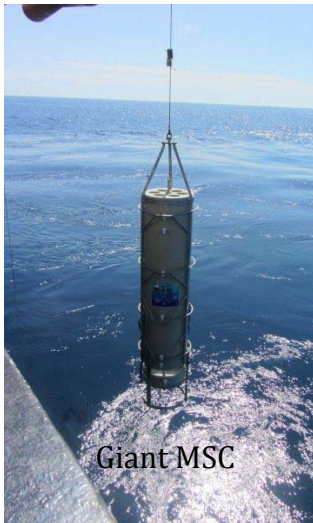
Table 1. Leucine sampling

Date	CTD	MSC	Depth	Bottle	Fraction	Comments
05/08/2014	2		20	17		
	2		45	11		
	2		60	8		
		1	20		Suspended, Slow and Fast (Giant Snowcatcher)	Above thermocline
		1	60		Suspended, Slow and Fast (Giant Snowcatcher)	Below thermocline
07/08/2014	6		15	21		
	6		35	17		
	6		50	9		
		5	50		Suspended, Slow and Fast (Giant Snowcatcher)	Below thermocline
		6	10		Suspended, Slow and Fast (Giant Snowcatcher)	Above thermocline
08/08/2014	9		20	19	T0,T6,T12,T18,T24	24hr calibration experiment
09/08/2014	10		50	10		
	10		38	13		
	10		10	21		
		10	50		Suspended, Slow and Fast (Giant Snowcatcher)	Below thermocline
		12	10		Suspended, Slow and Fast (Giant Snowcatcher)	Above thermocline
10/08/2014	14		92	3		Tidal sampling
	16		92	3		Tidal sampling
	18		91	11		Tidal sampling
	20		90	12		Tidal sampling
	22		95	8		Tidal sampling
	24		96	3		Tidal sampling
12/08/2014		31	100		Suspended, Slow and Fast (Small Snowcatcher)	Below thermocline dilution

						experiments
		32	100		Suspended, Slow and Fast (Small Snowcatcher)	Below thermocline dilution experiments

Marine Snow Catcher Deployments and Particle Characterization

Emma Cavan



Scientific Motivation

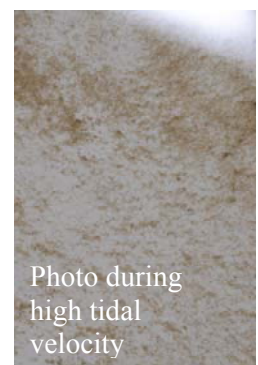
The marine snow catchers (MSCs) are an integral part of the Shelf Seas Biogeochemistry program. They are used to collect sinking particles in three sinking rate fractions, suspended, slow and fast sinking. On these particles bacterial production, aerobic respiration, sinking rates, organic chemistry and total mass can be measured. This method of capture allows *in situ* rates and states to be measured and links the pelagic and benthos, particularly during tidal studies. The MSCs allow us to quantify how much organic material produced in the surface either reaches the sediment, consumed in the water column or advected off the shelf.

Methods

At the three process stations (Candyfloss, Shelf break and Celtic Deep) we deployed the MSC above and below the deep chlorophyll maximum and above the sea bed (~20, 40 and 90 m respectively). The MSC is deployed with both ends open and a messenger fired to close the plungers and brought immediately to deck to be left to settle. Ideally it should stand for 2 hours in custom-made deck frames. After 2 hours the suspended fraction can be sampled, then this is drained and the slow sinking fraction is sampled. After which the tray with the fast sinking particles can be removed.

Deployments

Onboard we had the entire NOCS 'fleet' of MSCs. This consists of 3 small MSCs from the original design and 2 giant MSCs. The small MSCs can hold 100L water and the giant MSCs 350L. On this cruise we deployed 38 MSCs with a <90 % success rate in 8 days, a WORLD RECORD! During the 3 process station on this cruise the MSC was analyzed by all parties. During the tidal study the rate groups only analysed the CTD and so here organics, mass and microscope analysis were collected for. Additionally Michelle Barnett (UoS) undertook opportunistic sampling using the small MSC.



Limitations

In terms of deployments there are a few issues that arose with the giants MSCs on this cruise. Primarily we did not have the deck frames which meant lashing against the bulwark and using a step ladder to release the wire from the top of them, at 2.5 m tall this presents serious health and safety concerns. Also deploying them in any



high wind or sea state is risky due to their weight, $\sim 1/3$ of a ton. Additionally the clips used to secure the base to the top are not suitable for the size of the giant MSCs which resulted in using ratchet straps and as a result often leaking. All of the above led us to use the small MSCs when sampling in high resolution or high winds. This is a slight concern for future cruises as the small MSCs do not provide enough material for rate measurements. Even $1/4$ of the giant MSC tray in the summer may not have been enough for rate measurements.

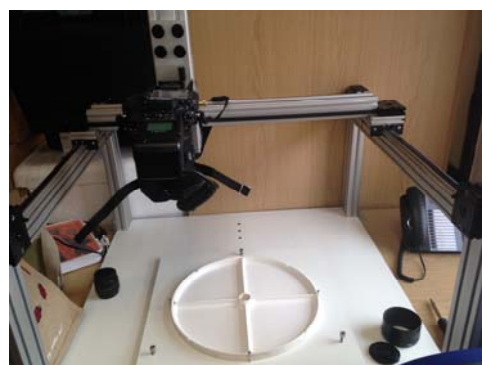
All rate measurements on this cruise needed the samples to remain in the dark from capture to measurements. This presented some trouble as the base of the MSCs are clear plastic. We used bin liners and tape to block out light. However when separating the top from the base the particles in the tray were exposed to light and therefore excitation and potential bleaching. Again we used a plastic bag to cover the base but some exposure is inevitable. Even using a clear tube for siphoning into a dark bottle exposes them to light. These things can and must be overcome by November cruise. The easiest solution is for particles to be collected at night when rate measurements are done.

We used $2/3$ of the small MSCs and during one of the final deployments part of the base of a small MSC cracked. Although still useable this shall need to be fixed before it can be used regularly at sea again.

Stephanie Wilson, Bangor University

Particle Characterization

To determine how representative the quarters of the tray of fast sinking particles are of the entire tray Stephanie Wilson built an imaging rig to photograph the trays before the samples were split and analysed by various groups. Depending on the size of the tray (small or giant) 30-100 photos were taken which can then be stitched together at a later date using Image J. When particles were collected for rate measurements these $1/4$ s had to be removed before flash photography could take place. Using the area of



particles and conversion rates of Alldredge (1998) particulate organic carbon content can be estimated. This is the conventional method used to estimate POC for the MSC fast sinking fraction. However as POC was also chemically measured the two methods can be compared.

The other motivation for the photography is to work out the proportion of faecal pellets compared to other particles in the tray such as aggregates.

When possible $1/4$ of a tray was also fixed in formalin. This is to allow later analysis in the lab using a microscope and further characterization of the type of particles in the Shelf Seas. In the November cruise Stephanie will continue with these measurements and also collect particles for molecular analysis with a focus on the role of zooplankton on export.

Table. Deployment Information

cruise	station	date	lat	long	event	Time	MSC	Type	depth	photo	formalin	organics	nitrification	bact prod	mass	notes
DY026	Candyfloss	05/08/2014	49 22.81	8 36.45	na	07:15	1	giant	20	3/4	1/4	1/2	0	1/4	na	process
DY026	Candyfloss	05/08/2014	49 22.81	8 36.45	na	08:30	2	giant	60	1/2	1/4	1/4	1/4	1/4	na	process
DY026	Candyfloss	05/08/2014	49 22.81	8 36.45	na	12:15	3	giant	100	1/2	1/4	1/2	0	1/4	na	process
DY026	Celtic deep	06/08/2014	48 2.32	9 43.6	na	14:10	4	small	35	1	0	0	0	0	na	micchelle
DY026	Shelf break	07/08/2014	48 34.19	9 30.61	na	06:45	5	giant	50	1/2	1/4	1/4	1/4	1/4	na	process
DY026	Shelf break	07/08/2014	48 34.19	9 30.61	na	11:50	6	giant	10	0	0	0	0	0	na	process
DY026	Shelf break	07/08/2014	48 34.19	9 30.61	na	13:40	7	giant	30	1	0	1	0	0	na	process
DY026	Celtic deep	08/08/2014	50 15.49	7 44.61	57	13:15	8	small	40	1	0	0	0	0	na	micchelle
DY026	Celtic deep	09/08/2014	50 15.49	7 44.61	59	06:45	9	giant	50	0	0	1/4	3/4	0	na	process
DY026	Celtic deep	09/08/2014	50 15.49	7 44.61	60	07:50	10	giant	50	0	0	1/4	0	3/4	na	process
DY026	Celtic deep	09/08/2014	50 15.49	7 44.61	63	08:15	11	small	50	1	3/4	1/4	0	0	na	process
DY026	Celtic deep	09/08/2014	50 15.49	7 44.61	70	11:30	12	giant	10	1/2	1/4	1/4	0	1/2	na	process
DY026	Celtic deep	09/08/2014	50 15.49	7 44.61	72	12:50	13	small	40	1	0	1/4	0	0	na	micchelle
DY026	Benthic	10/08/2014	51 9.39	6 34.3	87	08:45	14	small	90	1	1/4	1/2	0	0	1/4	Tidal study
DY026	Benthic	10/08/2014	51 9.39	6 34.3	89	09:45	15	small	90	1	1/4	1/2	0	0	1/4	Tidal study
DY026	Benthic	10/08/2014	51 9.39	6 34.3	91	10:45	16	small	90	1	1/4	1/2	0	0	1/4	Tidal study
DY026	Benthic	10/08/2014	51 9.39	6 34.3	93	11:45	17	small	90	1	1/4	1/2	0	0	1/4	Tidal study
DY026	Benthic	10/08/2014	51 9.39	6 34.3	95	12:45	18	small	90	1	1/4	1/2	0	0	1/4	Tidal study
DY026	Benthic	10/08/2014	51 9.39	6 34.3	97	13:45	19	small	90	1	1/4	1/2	0	0	1/4	Tidal study
DY026	Benthic	10/08/2014	51 9.39	6 34.3	99	14:45	20	small	90	1	1/4	1/2	0	0	1/4	Tidal study
DY026	Benthic	10/08/2014	51 9.39	6 34.3	101	15:45	21	small	90	1	1/4	1/4	0	0	1/2	Tidal study
DY026	Benthic	10/08/2014	51 9.39	6 34.3	103	16:45	22	small	90	1	1/4	1/2	0	0	1/4	Tidal study
DY026	Benthic	10/08/2014	51 9.39	6 34.3	105	17:45	23	small	90	1	1/4	1/2	0	0	1/4	Tidal study
DY026	Benthic	10/08/2014	51 9.39	6 34.3	107	18:45	24	small	90	1	1/4	1/2	0	0	1/4	Tidal study
DY026	Benthic	11/08/2014	51 12.3	6 07.45	114	12:50	25	small	38	0	0	0	0	0	na	leaked
DY026	Benthic	11/08/2014	51 12.3	6 07.45	115	13:00	26	small	20	0	0	0	0	0	na	leaked
DY026	Benthic	11/08/2014	51 12.3	6 07.45	118	16:00	27	small	40	1	0	0	0	0	na	micchelle
DY026	Benthic	11/08/2014	51 12.3	6 07.45	119	16:10	28	small	20	1	0	0	0	0	na	micchelle
DY026	Benthic	12/08/2014	51 8.11	6 37.77	133	06:10	29	small	100	0	0	0	0	0	na	baby clare test phosphate
DY026	Benthic	12/08/2014	51 8.11	6 37.77	134	06:30	30	small	100	1	1/2	0	0	0	na	steph
DY026	Benthic	12/08/2014	51 8.11	6 37.77	136	08:55	31	small	100	0	0	0	0	1	na	elena
DY026	Benthic	12/08/2014	51 8.11	6 37.77	137	09:10	32	small	100	0	0	0	0	1	na	elena
DY026	Benthic	12/08/2014	51 8.11	6 37.77	144	13:00	33	small	100	0	0	0	0	0	na	baby clare phosphate
DY026	Benthic	12/08/2014	51 8.11	6 37.77	145	13:10	34	small	100	0	0	0	0	0	na	no sample
DY026	Benthic	12/08/2014	51 8.11	6 37.77	147	15:45	35	small	30	1	0	0	0	0	na	micchelle
DY026	Benthic	12/08/2014	51 8.11	6 37.77	148	15:55	36	small	15	1	0	0	0	0	na	micchelle
DY026	Benthic	12/08/2014	51 8.11	6 37.77	151	20:00	37	small	30	1	0	0	0	0	na	micchelle
DY026	Benthic	12/08/2014	51 8.11	6 37.77	152	20:10	38	small	15	1	0	0	0	0	na	micchelle

Near Surface Gradients

Richard Sims, Plymouth Marine Laboratory, UK

Science Background

Physical and biological mechanisms affect the sea surface concentration and flux of carbon dioxide (CO₂) between the ocean and atmosphere. Gradients in temperature and salinity between the bulk sub-surface water and the sea/air interface are likely. Variations in phytoplankton primary production also have the potential to create vertical gradients in pCO₂. This studentship seeks to improve understanding of these physical and chemical gradients in near surface (<10 m) shelf waters.

This work will help answer a critical question within the Shelf Sea Biogeochemistry (SSB) research programme:

What are the current annual exchanges of carbon between UK/European shelf seas, the atmosphere, and the open ocean?

Cruise Objectives

- Test the deployment of the NSOP on board the ship to find a workable deployment strategy
- Obtain first data from the SSB cruises about near surface gradient
- Install IR-sensors on the front of the ship and get them continuously logging
- Familiarise myself with life at sea and with the Discovery so I can improve my setup for next time
- Help with the shelf sampling by collecting TA/DIC samples from the midday CTD
- Prepare and attach temperature sensors onto CEFAS SmartBuoy

Day by Day Account of Events

28th July to 1st August - setup days

Setup took much longer than expected mostly in part due to the installation required for the IR sensors on the bow of the ship. The IR sensors eventually started working once the 15core cable had been run up from the met lab up through to the met platform and into the logger box stored there. The accelerometer was not setup. Gas standards were stored in a rack in the hanger flowing into the deck lab where the remainder of my kit was stored. The PC, Nafion dryer, temperature sensor electronics, membrane equilibrator and peristaltic pump were located at the far end of the lab close to the sink with the bench to the left for working. The remainder of my equipment was kept underneath the bench except for the reel of tubing and the buckets on nylon rope. Nitrogen and compressed air cylinders put in the gas bottle store.

4th August - first deployment day

The lower IR sensor stopped working when we left Southampton and was patchy for the majority of the cruise, probably because of a loose connection. The NSOP winch faulty and would not work. Despite the winch not working, NSOP still deployed. We trailed deploying off the aft of the ship on the crane, this

proved to be unsuitable as the dp or movements by the bridge caused the buoy to rock, when there was a sharp movement by the ship the buoy almost flipped! It was eventually decided, with some hesitation from some NMF staff, to put it out of the CTD hanger on one of those winches with slack on the crane line it appeared to work well. It was noted that the CTD only logs at a maximum rate of once every 6 seconds, which is a minor issue. NSOP rope floats were ineffective at providing buoyancy.

5th August - repairs/transit

Lots of effort went into fixing the winch and discovered it was a wiring fault, fixed with the help of Jon Short.

6th August - first successful deployment/disaster

The IR sensor began working again which indicated that the problem was intermittent. NSOP deployed from the CTD crane unsuccessfully as even with a slightly slack cable NSOP was forced around considerably and eventually the strop was cut apart on the stainless steel plates. This was a blessing in disguise as the buoy happily drifted away from the ship and positioned by the two slack lines. This deployment last 3 hours, the lifting bar/strop hooked on to the crane using a pole during retrieval. An eyelet was welded onto the top of NSOP for the deployment the next day. The water column was being mixed heavily by the use of the aft thrusters, communication with the Captain to discuss this was poor on this day. The large head for the pump reduced its efficiency substantially.

7th August

NSOP was successfully deployed again today. The buoy was deployed on the crane at the back and lowered into the water using a quick release to detach it from the crane. The Captain agreed to turn off the DP after discussions with him. Slack lines were improved by making the bit near the buoy a chain so the nylon would not be damaged by the stainless plates. Pump flow rates are a problem as the waterside flow kept dropping during the deployment. Helped turn on and activate the Underway pCO₂ system on the *Discovery*.

8th August

Another successful deployment, the arm on the back of the ship extended the slack line point substantially and helped keep the buoy away. Bubbles seen on the equilibrator outflow indicative of non-equilibration probably due to a flow rate drop from 2L to 1L. Winch spool problem as the rope was not spooled by hand, this resulted in the Kevlar being sliced and the cage falling free, meaning it was recovered separately. GoPro footage of the shark taken on this day. Chata also got one discrete depth sample for her MIMS, this sample was not analysed. Liqui-Cel™ equilibrator cleaning in the evening.

9th August

Another successful deployment using a cleaned liquid-cel and a new piece of marprene tubing. No apparent flow rate problems.

10th August

Bad weather prevented deployments on this day instead I spent the day collecting CTD bottle water for Claire Mahaffy during the tidal cycle experiment using the Marine Snow Catchers.

11th August

Another successful deployment. Very small changes in pump efficiency. Considerable bad weather meant that NSOP shunted around a lot and the Captain was apprehensive of deploying NSOP in similar conditions again.

12th August

Temperature calibration in a water bath conducted in the morning. The deployment cut short as the battery enclosure lid was forced off by the surrounding stainless steel lifting it off. The enclosure then flooded with water and the circuit shorted and stopped working. NSOP redeployed and dried out. Re-fixed the Underway pCO₂ system on *Discovery* after the water error.

14th August

Final day of sampling and unfortunately the pumping efficiency dropped substantially in a very short amount of time. Less than 5 minutes to a flow less than 0.5L/min. At this point I decided to cancel the deployment and tried to fix the pump unsuccessfully.

Data Collected

- Sporadic Underway CO₂ measurements to compare with the on board system
- Primary Data- Vertical profiles of CO₂ (and ancillary information), temperature and salinity
- SST skin temperature and down welling irradiance
- 60 TA and DIC sample bottles for post analysis by NOC staff (not responsible for this)
- Underway pCO₂ system (not responsible for this)

Outcomes

- All of my objectives achieved to some extent on this cruise
- I had several successful deployments, I would have liked to have gotten more data as there were a lot of missed opportunities as a result of equipment not working in one form or another including winch failure, winch spool problems, pump problems and IR sensor wiring
- NSOP is not completely seaworthy and needs a lot of additional work before she is fit to come out to sea without any faults
- My relationship with the crew was good and this helped immensely during deployments
- I have not had an opportunity to connect the accelerometers up, which has meant I wasted a good opportunity to get data for that and test it

- The TA and DIC sampling whilst not difficult did cause some clashes as it demanded 40minutes of my time at moments when I was about to deploy
- Time was wasted on some of the CO₂ side of things, the gap between calibration and retrieval was also long at times
- A lot of time wasted trying to get the Underway CO₂ system working on the *Discovery*, this was not my responsibility but the data needed for comparisons
- I enjoyed my time at sea and I am now accustomed to life at sea and the ship

Success Rating of 7/10

Mesozooplankton Biomass and Metabolic Rates

Sarah Lou Carolin Giering

Scientific Motivation

Zooplankton play a significant role in the biogeochemical cycle of the sea as they ingest particulate organic matter and transform it into (1) CO₂ via respiration, (2) N-rich dissolved matter via excretion, and (3) particulate matter via the production of biomass, eggs and C-rich faecal pellets. The N-rich excretion products are likely to remain in the dissolved phase, whereas the C-rich faecal pellets may sink to depth at rates of up to 2700 m/d. This differential recycling, with N staying in the upper ocean and C exported to depth, has been postulated to enhance decoupling of C and N in shelf regions.

During DY026a, zooplankton biomass and composition was sampled using WP2 nets. These samples will compliment the SSB zooplankton biomass data time series by providing data for autumn. I further used this opportunity to trial the methods I proposed to do during the forthcoming SSB cruises. These experiments targeted measuring the process rates (excretion, sloppy feeding, and grazing) by mixed communities using incubation experiments.

Material & Methods

Abundance estimates

Eight WP2 nets fitted with non-filtering cod-ends and a closing mechanism were deployed at each process station: four during daytime and four during night-time to sample below and above the thermocline. Zooplankton of the size between 63-200 µm were collected using a 63-µm WP2 net hauled at 0.2 m/s. Zooplankton larger than 200 µm were collected using a 200-µm WP2 net hauled at 0.5 m/s. Collected zooplankton was size-fractionated into 63-200 µm, 200-500 µm, and >500 µm. Each size fraction split, half preserved in borax-buffered formaldehyde for identification and counts and half frozen at -80°C for CN analysis.

Rate-series experiments

The rate-series experiments aimed to measure different metabolic rates of the same 'mixed community'. To do so, I transferred the same group from one size class of zooplankton (63-200 µm and 200-500 µm) through sequential experiments determining rates of (1) excretion of DOC, ammonium, and nutrients, (2) sloppy feeding release of DOC, ammonium, and nutrients, and (3) ingestion. The order chosen to combine acclimation phases with actual rate measurements.

Sample summary

Fifty-one nets deployed in total (Table 1) at five stations. Two rate-series experiments, which measured excretion and sloppy release of DOC, ammonium, and nutrients, carried out. Ammonium and nutrient samples analysed onboard. DOC samples were frozen and stored at -20°C degrees for on-shore analysis.

Two grazing experiments carried out, from which one conducted in conjunction with a rate-series experiment. From the grazing experiments, samples taken for Chlorophyll and phytoplankton (preserved using Lugol's iodine).



Figure 1. Zooplankton abundance samples from 12th August. The colours indicate that the deep 63- μ m net collected sediments whilst the shallow 63- μ m net collected large phytoplankton from the chlorophyll maximum

Date	Daytime	Stn Name	Stn #	Net	Position (N) (W)		Time Open (hh:mm)	Time Closed (hh:mm)	Depth (m)	Mesh size (μm)	Use
04/08/14	Day	Test Station	005	1							
05/08/14	Day	Candyf loss	013	2	44°2.31	8°36.46	13:10	13:35	120-30	63	Frozen (2x)
05/08/14	Day	Candyf loss	014	3	44°2.31	8°36.46	13:41	14:10	120-30	63	Formalin
05/08/14	Day	Candyf loss	015	4	44°2.31	8°36.46	14:12	14:27	30-0	63	Frozen (2x)
05/08/14	Day	Candyf loss	016	5	44°2.31	8°36.46	14:29	14:42	30-0	63	Formalin
05/08/14	Day	Candyf loss	017	6	44°2.31	8°36.46	14:45	14:55	120-30	200	Frozen (2x)
05/08/14	Day	Candyf loss	018	7	44°2.31	8°36.46	14:59	15:10	120-30	200	Formalin
05/08/14	Day	Candyf loss	019	8	44°2.31	8°36.46	15:12	15:21	30-0	200	Frozen (2x)
05/08/14	Day	Candyf loss	020	9	44°2.31	8°36.46	15:22	15:30	30-0	200	Formalin
05/08/14	Night	Candyf loss	024	10	49°2.33	8°36.46	21:29	21:42	120-30	63	Frozen / Formalin
05/08/14	Night	Candyf loss	025	11	49°2.33	8°36.46	21:45	22:00	30-0	63	Frozen / Formalin
05/08/14	Night	Candyf loss	026	12	49°2.33	8°36.46	22:10	22:15	120-30	200	Frozen / Formalin
05/08/14	Night	Candyf loss	027	13	49°2.33	8°36.46	22:21	22:26	30-0	200	Formalin
05/08/14	Night	Candyf loss	028	14	49°2.33	8°36.46	22:27	22:30	30-0	200	Exp 1
07/08/14	Day	Shelf Break	036	15	48°3.421	9°30.64	07:08	07:16	120-50	63	Frozen / Formalin
07/08/14	Day	Shelf Break	037	16	48°3.421	9°30.64	07:27	07:37	50-0	63	Formalin

07/08		Shelf			48°3	9°30.			12		Frozen
/14	Day	Break	038	17	4.21	64	07:54	07:58	0-50	200	/
											Forma
											lin
											Frozen
											/
07/08		Shelf			48°3	9°30.			50-		Forma
/14	Day	Break	039	18	4.21	64	08:09	08:14	0	200	lin
											Frozen
											/
07/08		Shelf			48°3	9°35.			12		Forma
/14	Night	Break	050	19	4.74	3	20:50	21:01	0-50	63	lin
											Frozen
											/
07/08		Shelf			48°3	9°35.			50-		Forma
/14	Night	Break	051	20	4.74	3	21:13	21:18	0	63	lin
											Frozen
											/
07/08		Shelf			48°3	9°35.			12		Forma
/14	Night	Break	052	21	4.74	3	21:29	21:34	0-50	200	lin
											Frozen
											/
07/08		Shelf			48°3	9°35.			50-		Forma
/14	Night	Break	053	22	4.74	3	21:41	21:45	0	200	lin
											Frozen
											/
09/08		Celtic			51°0	6°35.			95-		Forma
/14	Day	Deep	064	23	8.27	12	07:35	07:42	50	63	lin
											Frozen
											/
09/08		Celtic			51°0	6°35.			50-		Forma
/14	Day	Deep	065	24	8.27	12	07:50	07:55	0	63	lin
											Frozen
											/
09/08		Celtic			51°0	6°35.			95-		Forma
/14	Day	Deep	066	25	8.27	12	08:20	08:27	50	200	lin
											Frozen
											/
09/08		Celtic			51°0	6°35.			50-		Forma
/14	Day	Deep	067	26	8.27	12	08:32	08:40	0	200	lin
09/08		Celtic			51°0	6°35.			50-		Forma
/14	Day	Deep	068	27	8.27	12	08:43	08:48	0	200	lin
											Exp 2
09/08		Celtic			51°0	6°38.			25-		Forma
/14	Night	Deep	080	28	8.84	49	20:38	20:42	0	200	lin
											Exp 3
											Frozen
											/
09/08		Celtic			51°0	6°38.			95-		Forma
/14	Night	Deep	081	29	8.84	49	20:47	20:59	50	200	lin
09/08		Celtic			51°0	6°38.			50-		misfire
/14	Night	Deep	082	30	8.84	49	21:06	21:09	0	200	d
09/08		Celtic			51°0	6°38.			50-		misfire
/14	Night	Deep	083	31	8.84	49	21:15	21:20	0	200	d
09/08		Celtic			51°0	6°38.			50-		misfire
/14	Night	Deep	084	32	8.84	49	21:22	21:29	0	200	d

09/08	Night	Celtic Deep	085	33	51°08.84	6°38.49	21:35	21:38	50-0	200	Frozen / Formalin
11/08	Day	Benthic A	116	34	51°12.7	6°08.49	12:34	12:51	40-0	63	Exp 4
11/08	Day	Benthic A	120	35	51°12.08	6°07.96	15:40	15:48	90-40	63	Discarded Frozen / Formalin
11/08	Day	Benthic A	121	36	51°12.08	6°07.96	16:00	16:10	90-50	63	Frozen / Formalin
11/08	Day	Benthic A	122	37	51°12.08	6°07.96	16:16	16:22	50-0	63	Frozen / Formalin
11/08	Day	Benthic A	123	38	51°12.08	6°07.96	16:33	16:40	90-50	200	Frozen / Formalin
11/08	Day	Benthic A	124	39	51°12.08	6°07.96	16:47	16:51	50-0	200	Frozen / Formalin
11/08	Night	Benthic A	125	40	51°11.94	6°05.76	20:35	20:41	90-50	63	Frozen / Formalin
11/08	Night	Benthic A	126/7	41	51°11.94	6°05.76	20:54	21:02	50-0	63	Frozen / Formalin
11/08	Night	Benthic A	128	42	51°11.94	6°05.76	21:08	21:17	90-50	200	Frozen / Formalin
11/08	Night	Benthic A	129	43	51°11.94	6°05.76	21:26	21:30	50-0	200	Frozen / Formalin
12/08	Day	Celtic Deep	138	44	51°08.91	6°36.24	08:49	08:57	10-0-40	63	Frozen / Formalin
12/08	Day	Celtic Deep	139	45	51°08.91	6°36.24	09:07	09:14	40-0	63	Frozen / Formalin
12/08	Day	Celtic Deep	140	46	51°08.91	6°36.24	09:27	09:32	10-0-40	200	Frozen / Formalin

12/08 /14	Day	Celtic Deep	141	47	51°0 8.91	6°36. 24	09:43	09:48	40- 0	200	Frozen / Forma lin
12/08 /14	Night	Celtic Deep	143	48	51°0 8.1	6°37. 89	20:40	20:49	10 0- 40	63	Frozen / Forma lin
12/08 /14	Night	Celtic Deep	144/a	49	51°0 8.1	6°37. 89	20:58	21:04	40- 0	63	Frozen / Forma lin
12/08 /14	Night	Celtic Deep	144/ b	50	51°0 8.1	6°37. 89	21:12	21:17	10 0- 40	200	Frozen / Forma lin
12/08 /14	Night	Celtic Deep	145	51	51°0 8.1	6°37. 89	21:25	21:30	40- 0	200	Frozen / Forma lin

Sediment Cores

Matthew Bone, University of East Anglia, UK

Departed National Oceanography Centre Southampton, 14:00 hours on 3rd August 2014.

Heading out of Southampton Docks as the cumulus clouds rolled over the mask of the ship greeting the English Channel with a roar. Our first meeting held soon after giving the chance to meet all the Scientists aboard and briefly explain what each member intends to carry out.

My contribution on this cruise would be to collect sediment from the designated sites via a NIOZ sediment core, and look at various parameters associated with the collected sample.

The main experiment aboard would examine the change in ammonium (NH_4^+) as a sheer and vertical stress is applied to the sediment core. This would be carried out by placing a sediment erosion device (FloWave) directly into the core. The experiment would run continuously for approximately two hours and the NH_4^+ analysed at a high time resolution (139 times per second) using a hacked High Performance Liquid Chromatograph (HPLC). This set up was installed and ensured was in working order before the cruise set out. On 3rd August a plan was agreed that would accommodate the experimental needs as well as the crew. A preliminary experimental protocol to undertake a multitude of experiments was also drawn up:

- Sub sampling the mud and freezing for microbiological work
- Subsampling and incubating mud in the bottom water
- Running the resuspension experiments and measuring NH_4^+
- Sampling the resuspension experiments for $^{15}\text{N}/^{18}\text{O}$ isotopes
- Setting up a way to continuously measure the concentration on NH_4^+ in the surface water using the underway system

On 4th August several trials were planned to ensure the working of deployable equipment. In this time, there was no planned deployment of the sediment corer and time spent creating a method to sample from the underway system. The first trial of this method was successfully carried out at a stationary site (49 48.80412N, 5 28.65816W) from 12:33 BST onwards.

A second continuous sampling was carried out during transit from the stationary location (49 47.24790N, 5 41. 556566W) to the CaNDyFloSS site. The continuous sampling continued for 640 minutes until the experiment stopped as the pressure exceeding the maximum limit. The

pressure fluctuation caused problems during analysis; as the pressure varied by 10bars, the lumosity changed accordingly. The route of the problem is unknown; however, pressure limits can be applied to control the minimum and maximum limits. A disadvantage with setting limits is, if they are exceeded, there is a failsafe shutdown of the machine.

During the evening of 4th August continuous checking of the sampling was undertaken throughout the night to ensure an effective working method.

4th August 2014

A new system was set up to continuously measure from the underway system using the HLPC. Two attempts were made and a calibration of the machine carried out (Lat 49 47.24790, Lon 5 41.556566).

A detailed fluorescent scan was undertaken on the surface seawater from the under way system to determine the most suited wavelength to measure ammonium using the HPLC set up.

5th August 2014

Two cores were taken from the Shelf break (Lat 49 22.33440, Lon 8 36.46206), but due to the neoprene not stuck on effectively, both cores slumped and leaked water thus rendering them useless for resuspension analysis due to chemical change.

Four incubations were carried instead taking syringe cores from three depths and one throughout the depths sampled and spiked with a nitrification inhibitor. The experiments ran for the next six hours to determine any rapid change in ammonium.

6th August 2014

A calibration was carried out on the HPLC to analyse nano-molar concentrations of ammonium.

7th August 2014

Three cores were taken between 10:00 & 12:00 from Benthic Site 'H' (Lat 48 34.58532, Lon 9 30.96324). The first core slumped during transit on deck, with water and mud leaking out. The overlying water was syphoned off into brown bottles, and the sediment subsampled using syringes. The sediment was added to the bottles. The incubation experiments ran for 6hours. Due to software error, the data from these experiments were lost. The second core was used in a FloWave resuspension device experiment. A second high-resolution scan was carried out from the overlying water on the NIOZ core before the FloWave experiment. Samples were taken for microbiological work – frozen at -80°C.

9th August 2014

Three cores (Lat 51 7.09974, Lon 6 37.49844).

ATU experiments undertaken from sediment collected at Benthic Site 'H', Core one.

Incubation experiments measuring ammonium undertaken from sediment collected at Benthic Site 'H' Core one.

Samples were taken for microbiological work – frozen at -80°C.

A FloWave resuspension experiment was ran on intact Core three measuring ammonium, Core one.

The HPLC set to measure continuously from the underway. On several occasions it failed, but was up and running to measure the impact of increased turbulence and wind forcing upon ammonium concentrations in the ocean.

10th August 2014

A FloWave experiment undertaken from a Core taken at 11:00 from Benthic Site 'H'.

Sampling continuously from the underway system.

11th August 2014

Three mud cores taken from Benthic Site 'A'.

Samples were taken for microbiological work – frozen at -80°C.

Incubation experiments set up to measure nitrification of NH₄⁺ within the sediment.

- Control
- Control + sediment
- Control + sediment + inhibitor

Underway measurements made during the Spring tide.

12th August 2014

A FloWave experiment was undertaken.

13th August 2014

A NIOZ sediment core was taken from Benthic Site 'A' at 07:30 Lat 51 12.56622, Lon 6 3.75258). A sediment resuspension experiment then ran for the next hour with measurements of oxygen, nutrients, DOC, 15N/18O and NH₄⁺ taken throughout.

Resuspension experiment on Core 9 taken from Benthic site 'A'.

Microbial samples taken from Core 6 and frozen at -80°C.

Calibrated machine.

Cruise Report DY026b

CRUISE SUMMARY REPORT

FOR COLLATING CENTRE USE

Centre: BODC Ref. No.:

Is data exchange

restricted Yes In part
No

SHIP enter the full name and international radio call sign of the ship from which the data were collected, and indicate the type of ship, for example, research ship; ship of opportunity, naval survey vessel, etc.
enter the unique number, name or acronym assigned to the cruise (or cruise leg, if appropriate).

Name: RRS Discovery

Call Sign: 2FGX5

Type of ship: Research Vessel

CRUISE NO. / NAME DY026b

CRUISE PERIOD start 16/08/2014 to 24/08/2014 end
(set sail) day/ month/ year day/ month/ year (return to port)

PORT OF DEPARTURE (enter name and country) Southampton, UK

PORT OF RETURN (enter name and country) Southampton, UK

RESPONSIBLE LABORATORY enter name and address of the laboratory responsible for coordinating the scientific planning of the cruise

Name: Cefas

Address: Lowestoft Laboratory, Pakefield Road, Lowestoft, Suffolk. NR33 0HT

Country: UK

CHIEF SCIENTIST(S) enter name and laboratory of the person(s) in charge of the scientific work (chief of mission) during the cruise.

Dave Sivyer, Cefas. (as above) dave.sivyer@cefas.co.uk

OBJECTIVES AND BRIEF NARRATIVE OF CRUISE enter sufficient information about the purpose and nature of the cruise so as to provide the context in which the report data were collected.

DY026 was funded as part of NERC's Shelf Sea Biogeochemistry (SSB) programme. This programme aims to reduce uncertainty in our process understanding of the cycling of nutrients and carbon, and the controls on primary and secondary production in both the UK and European shelf seas, and in wider global biogeochemical cycles. A series of long-term moorings and gliders were deployed in the Celtic Sea in early 2014 (DY008) and will remain in the water until late summer 2015. They will provide an unprecedented record of both physical and biogeochemical measurements across a full seasonal cycle providing the research community with (a) a long term record of the parameters controlling biogeochemical cycling rates and pathways, (b) a background against which to set process studies carried out on subsequent cruises and (c) essential data for model validation and development.

The main objective of DY026b was to service the moorings. Specifically:

- (a) to service 8 moorings/landers distributed across 5 different sites in the Celtic Sea
- (b) to calibrate the moorings

Moorings calibration achieved via CTD casts pre-recovery and post-deployment. Samples were collected for DIC/TA, DIN, DON, Chl-a, SPM and oxygen. The shipboard CTD winch failed for a part of the cruise so the Cefas ESM2 profiler deployed instead with water samples collected from the clean lab supply.

Additional CTDs performed in between the main study sites to build a more complete cross-shelf picture of key biogeochemical and physical gradients (names A1-A5).

Most moorings were deployed on JC105 and were due to be serviced again on CEND22/14 and DY018

PROJECT (IF APPLICABLE) if the cruise is designated as part of a larger scale cooperative project (or expedition), then enter the name of the project, and of organisation responsible for co-ordinating the project.

Project name: Shelf Sea Biogeochemistry

Coordinating body: NERC

PRINCIPAL INVESTIGATORS: Enter the name and address of the Principal Investigators responsible for the data collected on the cruise and who may be contacted for further information about the data. (The letter assigned below against each Principal Investigator is used on pages 2 and 3, under the column heading ‘PI’, to identify the data sets for which he/she is responsible)

- A. Jo Hopkins. NOC, Liverpool**
- B. Dave Sivyver. Cefas**
- C. Claire Mahaffey. University of Liverpool**
- D. Sue Hartman. NOC, Southampton**
- E. Malcolm Woodward. PML**
- F. Alex Poulton. NOC, Southmapton**
- G. Matthew Palmer. NOC, Liverpool**
- H. Peter Statham. University of Southampton**
- I. Alex Souza. NOC, Liverpool**
- J. James Fox, University of Essex**
- K. Tom Rippeth, University of Bangor**
- L. Kieran Curran, Plymouth Marine Laboratory, kic@pml.ac.uk**
- M. Sam Ward, National Oceanography Centre, sjw@noc.ac.uk**
- N. David White, MARS**
- O. Stuart Painter, NOC, Southampton**

MOORINGS, BOTTOM MOUNTED GEAR AND DRIFTING SYSTEMS

This section should be used for reporting moorings, bottom mounted gear and drifting systems (both surface and deep) deployed and/or recovered during the cruise. Separate entries should be made for each location (only deployment positions need be given for drifting systems). This section may also be used to report data collected at fixed locations which are returned to routinely in order to construct ‘long time series’.

PI	APPROXIMATE POSITION						DATA TYPE	DESCRIPTION
	LATITUDE			LONGITUDE				
See top of page .	deg	min	N/S	deg	min	E/W	enter code(s) from list on last page.	Identify, as appropriate, the nature of the instrumentation the parameters (to be) measured, the number of instruments and their depths, whether deployed and/or recovered, dates of deployments and/or recovery, and any identifiers given to the site.
	49	24	N	8	36	W		SITE 1
A							H72	23 temperature and conductivity loggers (3 x SBE 16 +, 4 x

							SBE 37, 16 x mini temperature loggers): recovered 21/08/2014, deployed 22/08/2014
A							D90 Bedframe with SBE 16+ (temperature, pressure, conductivity). Recovered 19/06/2014, deployed 22/06/2014
A							D71 150 kHz ADCP mounted in bedframe (upward looking). Recovered 22/08/2014, deployed 22/08/2014
L							D71 600 kHz ADCP mounted in bedframe (upward looking). Recovered 22/08/2014, deployed 22/08/2014
L							D71 3 x 600 kHz ADCPs mounted inline on mooring wire. Recovered 21/08/2014, deployed 22/08/2014.
B							H90/H1 6/M02/ B02/H2 1 Cefas SmartBuoy with temperature and conductivity logger (Aanderaa & SBE37), optical backscatter (Seapoint), fluorometer (Seapoint), oxygen sensor (Aanderaa). All approx. 1-2 m below surface. PAR sensor in air (Licor). Recovered 21/08/2014. Deployed 21/08/2014.
B							H24/H2 5/H26/ H22 Cefas SmartBuoy mounted 50 port water sampler, 1 m below surface. Recovered 21/08/2014. Deployed 21/06/2014
	50	36	N	7	2	W	SITE 2
B							D90/H1 6/B02/ H21 Cefas minilander with temperature and conductivity (Aanderaa), optical backscatter (Seapoint), fluorometer (Seapoint), oxygen sensor (Aanderaa). Recovered, 19/08/2014, deployed 20/08/2014.
J							D71 600 kHz ADCP mounted in minilander (upward looking). Recovered, 19/08/2014, deployed 20/08/2014.
	51	3	N	6	36	W	SITE 3
B							D90/H1 6/B02/ H21 Cefas minilander with temperature and conductivity (Aanderaa), optical backscatter (Seapoint), fluorometer (Seapoint), oxygen sensor (Aanderaa). Not recovered, no new instruments deployed.
J							D71 600 kHz ADCP mounted in minilander (upward looking). Not recovered.
	51	7	N	6	10	W	SITE 5
B							D90/H1 6/B02/ H21 Cefas minilander with temperature and conductivity (Aanderaa), optical backscatter (Seapoint), fluorometer (Seapoint), oxygen sensor (Aanderaa). Not recovered.
J							D71 600 kHz ADCP mounted in minilander (upward looking). Not recovered
	51	8	N	6	34	W	SITE 4
B							H90/H1 6/M02/ B02/H2 1 Cefas SmartBuoy with temperature and conductivity logger (Aanderaa), optical backscatter (Seapoint), fluorometer (Seapoint), oxygen sensor (Aanderaa). All approx. 1-2 m below surface. PAR sensor in air (Licor). Deployed 17/08/2014.
J							D71 300 kHz upward looking ADCP mounted inline at approx. 80 m below surface. Deployed 17/08/2014.
J							H72 Temperature loggers (SBE56 and SBE39) mounted at 10, 20, 30, 40, 60 and 80 m below surface. Deployed 17/08/2014.

B						H24/H25/H26/H22	Cefas SmartBuoy mounted 50 port water sampler, 1 m below surface. Deployed 17/08/2014.
							Please continue on separate sheet if necessary

SUMMARY OF MEASUREMENTS AND SAMPLES TAKEN

Except for the data already described on page 2 under ‘Mooring, Bottom Mounted Gear and Drifting Systems’, this section should include a summary of all data collected on the cruise, whether they be measurements (e.g. temperature, salinity values) or samples (e.g. cores, net hauls).

Separate entries should be made for each distinct and coherent set of measurements or samples. Different modes of data collection (e.g. vertical profiles as opposed to underway measurements) should be clearly distinguished, as should measurements/sampling techniques that imply distinctly different accuracy’s or spatial/temporal resolutions. Thus, for example, separate entries would be created for i) BT drops, ii) water bottle stations, iii) CTD casts, iv) towed CTD, v) towed undulating CTD profiler, vi) surface water intake measurements, etc.

Each data set entry should start on a new line – it’s description may extend over several lines if necessary.

NO, UNITS : for each data set, enter the estimated amount of data collected expressed in terms of the number of ‘stations’; miles’ of track; ‘days’ of recording; ‘cores’ taken; net ‘hauls’; balloon ‘ascents’; or whatever unit is most appropriate to the data. The amount should be entered under ‘NO’ and the counting unit should be identified in plain text under ‘UNITS’.

PI	NO	UNITS	DATA TYPE	DESCRIPTION
see page 2	see above	see above	Enter code(s) from list on last page	Identify, as appropriate, the nature of the data and of the instrumentation/sampling gear and list the parameters measured. Include any supplementary information that may be appropriate, e.g. vertical or horizontal profiles, depth horizons, continuous recording or discrete samples, etc. For samples taken for later analysis on shore, an indication should be given of the type of analysis planned, i.e. the purpose for which the samples were taken.
A	7	profiles	D90/H21/	CTD profiles (temperature, salinity, pressure, oxygen, beam transmission,

			H16/ B02/ H17	fluorescence, PAR/Irradiance, light scatter)
F	21	samples	B02	Filtration of samples for laboratory analysis of chlorophyll-a (from CTD niskin bottles)
G	21	samples	P01	Filtration of samples for laboratory analysis of suspended particulate matter (from CTD niskin bottles)
B	21	samples	H21	Dissolved oxygen samples from CTD niskin bottles
E	21	samples	H75/ H25/ H26/ H22	Collection of dissolved inorganic nutrient samples for laboratory analysis (nitrate+nitrite, nitrite, silicate, phosphate) (from CTD niskin bottles)
C	21	samples	B06	Collection of dissolved organic matter (DOC, DON, DOP) samples for laboratory analysis (from CTD niskin bottles)
D	21	samples	H90/ H27	Collection of dissolved inorganic carbon and total alkalinity samples for laboratory analysis (from CTD niskin bottles)
E	2	samples	H75/ H25/ H26/ H22	Collection of dissolved inorganic nutrient samples from underway non-toxic supply for laboratory analysis (nitrate+nitrite, nitrite, silicate, phosphate)
C	2	samples	B06	Collection of dissolved organic matter (DOC, DON, DOP) samples from underway non-toxic supply for laboratory analysis
D	2	samples	H90/ H27	Collection of dissolved inorganic carbon and total alkalinity samples from underway non-toxic supply for laboratory analysis
N	9	days	H74	Underway PCO2 analysis
A	2	profiles	H21/ B02/ H17/P 01	CTD profiles (temperature, salinity, pressure, oxygen, optical back scatter (sediment load), chlorophyll fluorescence, PAR/Irradiance)
Please continue on separate sheet if necessary				

TRACK CHART: You are strongly encouraged to submit, with the completed report, an annotated track chart illustrating the route followed and the points where measurements were taken.

Insert a tick(✓) in this box if a track chart is supplied

X



Approximate cruise track

GENERAL OCEAN AREA(S): Enter the names of the oceans and/or seas in which data were collected during the cruise – please use commonly recognised names (see, for example, International Hydrographic Bureau Special Publication No. 23, ‘Limits of Oceans and Seas’).

NE Atlantic

GEOGRAPHIC COVERAGE - INSERT 'X' IN EACH SQUARE IN WHICH DATA WERE COLLECTED

